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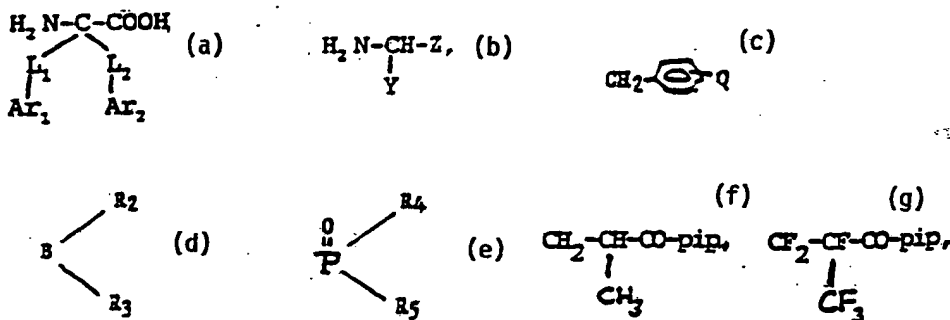
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(54) Title: INHIBITORS AND SUBSTRATES OF THROMBIN**(57) Abstract**

Peptides which act as inhibitors or substrates of thrombin are derived from the formula: D-Phe-Pro-Arg or its analogues, wherein Phe is substituted by (a), wherein Ar₁ and Ar₂ are the same or different and are selected from the group consisting of phenyl, thienyl, pyridyl, naphthyl, thionaphthyl, indolyl and saturated groups corresponding to these, optionally substituted by up to three groups selected from C₁-C₃ alkyl and C₁-C₃ alkoxy, L₁ and L₂ are the same or different and are selected from the group consisting of CH₂, CH₂-CH₂, O-CH₂, S-CH₂, Ar-L taken together optionally means H, diphenyl-methyl, fluorenyl or saturated groups corresponding to these, but one of the Ar-L cannot be H when the other Ar-L means H or benzyl, and Arg may be substituted by an amino acid of formula (b), wherein Y = [CH₂]_n-Q, (c), where Q = H, amino, amidino, imidazole, guanidino or isothioureido and n = 1-5, preferably 3-5, or C₃-C₉ alkyl and C₇-C₁₀ aryl or alkylaryl optionally substituted by up to three groups selected from hydroxy and C₁-C₄ alkoxy; Z = CN, COR₁, (d) or (e), where R₁ = H, OH, CH₂Cl, CH₂-CH₂-CO-pip, CF₂-CF₂-CO-pip, (f), (g), CH₂-CH₂-CO-Pro-NHEt, CF₂-CF₂-CO-Pro-NHEt or a chromophoric group e.g. pNA, MCA, R₂ and R₃ are the same or different and are selected from the group consisting of OH, OR₆ and NR₆R₇, or R₂ and R₃ taken together represent the residue of a diol; where R₆ and R₇, which are the same or different, are C₁-C₁₀ alkyl, phenyl or C₆-C₁₀ arylalkyl, R₄ and R₅ are the same or different and are selected from R₂, R₃, Gly-pip, Ala-pip or Gly-Pro-NHEt.

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INHIBITORS AND SUBSTRATES OF THROMBIN

This invention relates to thrombin inhibitors and substrates.

Thrombin, the last enzyme in the coagulation system, cleaves soluble fibrinogen to fibrin, which is then crosslinked and forms an insoluble gel forming the matrix for a thrombus. When a vessel is damaged, the above process is necessary to stop bleeding. Under normal circumstances there is no measurable amount of thrombin present in plasma. Increase of the thrombin concentration can result in formation of clots, which can lead to thromboembolic disease, one of the most common serious medical problems of our time.

Thrombin contributes to haemostatic control by means of several biological reactions. In addition to its primary function, the conversion of fibrinogen to fibrin, thrombin activates Factor XIII, which is responsible for the crosslinking of fibrin. Thrombin also acts by means of a positive feedback mechanism involving the activation of Factors V and VIII, which both are necessary for its own formation from prothrombin. Thrombin has another essential role: its binding to platelets initiates platelet release and aggregation which is responsible for primary haemostasis.

Fibrinolysis is the process which causes an enzymatic dissolution of fibrinogen and fibrin clots. Plasma contains a protein, plasminogen, which under the influence of various activators is converted to plasmin, a proteolytic enzyme, the activity of which resembles that of fibrin. Plasmin breaks down fibrin to fibrin degradation products.

Under normal conditions, the fibrinolysis system is in balance with the coagulation system. Small thrombi formed in the blood stream can be dissolved enzymatically and the circulation through the vessels can be restored by the

activation of the fibrinolytic system in the body. If the fibrinolytic activity is too high, it may cause or prolong bleeding and if it is too low compared to the activity of the coagulation system, there is a risk of thrombosis.

The reactions of thrombin are further controlled by natural inhibitors in plasma. The most important of these are antithrombin III and heparin. These two compounds have been isolated and are therapeutically and prophylactically used in conditions where there is an imbalance in the haemostatic mechanisms with risk for prothrombin activation.

Mainly two types of therapeutic agents are used for the prevention of thrombosis. The heparins act by accelerating the inhibition of thrombin by antithrombin III. Coumarin derivatives, the oral anticoagulants, e.g. Warfarin, prevent the generation of thrombin by blocking the post-translational vitamin K-dependent γ -carboxylation in the synthesis of prothrombin. Neither Heparin nor Warfarin are ideal. Heparin must be given parenterally and as it functions as a cofactor to antithrombin III it has no effect without this inhibitor. The effect of Warfarin develops very slowly and individual doses must be adjusted by frequent tests. None of these anticoagulants is specific for thrombin, they also inhibit other serine proteases and both of them may cause bleeding if the doses are not correctly balanced.

Thus, direct acting, specific thrombin inhibitors, having oral activity would be useful alternatives to the above anticoagulants. Much research in this area has resulted in the synthesis of different kinds of inhibitors of thrombin.

By imitating amino acid sequences of fibrinogen, the important natural substrate of thrombin, several good short peptide substrates for thrombin have been synthesized. The very first developed sequence with affinity for the active site of thrombin was Phe-Val-Arg [1] which mimics the fibrinogen sequence preceding the bond split by thrombin. This sequence has later been

improved to give D-Phe-Pro-Arg and D-Phe-Pip-Arg which have been used in chromogenic substrates, e.g. D-Phe-Pro-Arg-pNA and D-Phe-Pip-Arg-pNA [1] and in inhibitors of thrombin, e.g. the peptide aldehyde D-Phe-Pro-Arg-H [2], the irreversible inhibitor D-Phe-Pro-Arg-CH₂Cl [3], inhibitors with a ketomethylene bond e.g. D-Phe-Pro-Arg-k-Gly-piperidide [4] and in the recently synthesized peptide boronic acid inhibitors e.g. Z-D-Phe-Pro-boroArg [5] and the nitrile: Boc-D-Phe-Pro-ArgCN [6].

Thus, D-Phe-Pro-Arg has been considered the best sequence for about 15 years, and it has been shown to have very good affinity for the active site of thrombin, in substrates (K_m around $10^{-6}M$) as well as in inhibitors (K_i $10^{-7}M$ to $10^{-9}M$).

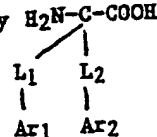
We have now found that by exchanging Phe, in the D-Phe-Pro-Arg sequence, for some unnatural, aromatic amino acids, with a specified structure, and by using these new sequences to construct novel substrates and inhibitors we obtained significantly improved substrate and inhibitor properties. The new substrates show better kinetic constants (K_m and k_{cat}) and the inhibitors better inhibition constant (K_i).

Reduction of blood pressure is a side effect observed in many of the previous thrombin inhibitors containing Arg or Arg analogues like Gpa and Apa [7]. This side effect which in some compounds can be disturbingly serious is believed to depend on the positively charged guanidino or amidino group of the side chain of Arg or its analogues. Surprisingly, this side effect of inhibitors in the present application is markedly reduced even when the inhibitors have an Arg or Arg analogue.

We have also surprisingly found that by changing the side chain to a non-basic alkyl or alkylaryl group of a certain size, the affinity for thrombin is still very good although the affinity for other serine proteases is greatly reduced, i.e. these inhibitors/substrates are more specific for thrombin than corresponding compounds containing Arg. With this non-basic side chain the

blood pressure lowering side effect is greatly reduced.

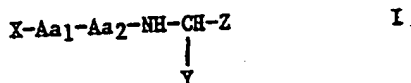
The present invention provides thrombin inhibitors and substrates derived from D-Phe-Pro-Arg or its analogues wherein Phe is substituted by $\text{H}_2\text{N}-\text{C}-\text{COOH}$




and Arg may be substituted by $\text{H}_2\text{N}-\text{CH}-\text{COOH}$.

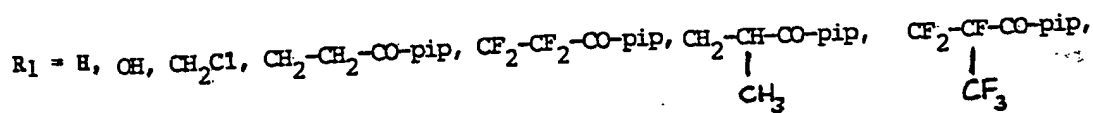
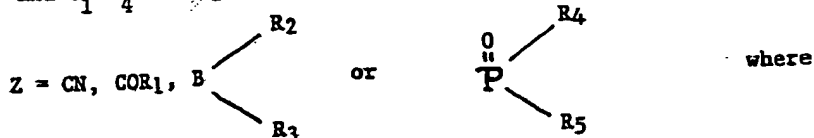


Suitably, the inhibitors/substrates are of formula I, in which



X = H, CH₃ or an N-protecting group, e.g. Ac, Bz, Cbz, Boc;

Y = [CH₂]_n-Q, CH₂--Q where Q = H, amino, amidino, imidazole, guanidino or isothioureido and n = 1-5, preferably 3-5, or C₃-C₉ alkyl and C₅-C₁₀ aryl or alkylaryl optionally substituted by up to three groups selected from hydroxy and C₁-C₄ alkoxy;

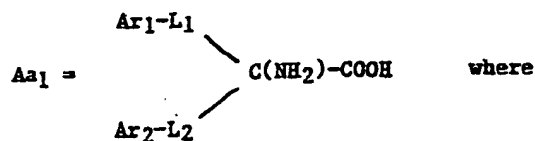


CH₂-CH₂-CO-Pro-NHET, CF₂-CF₂-CO-Pro-NHET or a chromophoric group e.g. pNA, MCA,

R₂ and R₃ may be the same or different and are selected from the group consisting of OH, OR₆ and NR₆R₇, or R₂ and R₃ taken together represent the residue of a diol; where R₆ and R₇, which may be the same or different, are C₁-C₁₀ alkyl, phenyl or C₆-C₁₀ arylalkyl,

R₄ and R₅ may be the same or different and are selected from R₂, R₃,

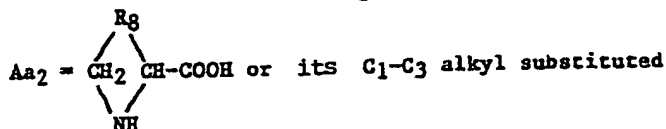
Gly-pip, Ala-pip or Gly-Pro-NHET;



Ar₁ and Ar₂ may be the same or different and are selected from the group consisting of phenyl, thienyl, pyridyl, naphthyl, thionaphthyl, indolyl and saturated groups corresponding to these, optionally substituted by up to three groups selected from C₁-C₃ alkyl and C₁-C₃ alkoxy,

L₁ and L₂ may be the same or different and are selected from the group consisting of CH₂, CH₂-CH₂, O-CH₂, S-CH₂;

Ar-L taken together may mean H, diphenyl-methyl, fluorenyl or saturated groups corresponding to these, but one of the Ar-L cannot be H when the other Ar-L means H or benzyl;



derivatives, where R₈ = CH₂, CH₂-CH₂, S-CH₂, S-C(CH₃)₂ or CH₂-CH₂-CH₂.

Preferably the Phe substitute is Dpa, Nal or Dbg and preferable Arg substitute includes Irg, Gpa, Apa and non-basic amino acids such as Pgl, Mbg, Chg.

Examples of compounds which may be preferably used in the invention include:

Ac-D-βNal-Pro-boroArg pinanediol ester

Z-D-Dpa-Pro-boroIrg pinanediol ester

Z-D-Dpa-Pro-boroPgl pinacol ester

Ac-D-βNal-Pro-boroMbg pinanediol ester

CH₃-D-Dpa-Pro-Arg-H

Boc-D-Dpa-Pro-Gpa-H

CH₃-D-Dpa-Thi-Mbg-H

H-D-Dpa-Pro-Arg-k-Gly-pip

Z-D-Dpa-Pro-Arg-CH₂Cl
Boc-D-Dpa-Pro-ArgCN
H-Dpa-Pro-Arg^P (OPh)₂
H-D-SNal-Pro-Pgl^P (OPh)-Gly-pip
H-D-Dpa-Pip-Arg-pNA
H-D-SNal-Pro-Chg-pNA

Further examples of compounds which may be preferably used in the invention are those listed in Examples 10 to 22 below.

Inhibition data for some of the new compounds are shown in Tables 1-7. The advantages of replacing Phe by amino acids according to the invention are clearly shown in the K_i values, which are generally 3 to 10 times better for the new compounds, as well as in the prolongation of the thrombin time. The importance of the D-form of the N-terminal amino acid is also evident from Table 1. The drastic reduction of the blood pressure lowering side-effect with compounds according to the invention is shown in Table 2.

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TABLE 1 - In-vitroassays

		Ki (μM)	TT (μM)	APTT (μM)
	D-Phe-Pro-Gpa-k--Gly-Pip	1.3	4.9	
4	D, L-Dpa-Pro-Arg--k-Gly-Pip	0.6		45.0
6	D-Dpa-Pro-Arg--k-Gly-Pip	0.2	0.65	8.0
7	L, Dpa-Pro-Arg--k-Gly-Pip	1.7	7.5	100.0
8	D-Fgl-Pro-Arg--k--Gly-Pip		50.0	
9	D, L- Q -Nal-Pro-Arg--k-Gly-Pip	13.5		
14	D, L-8-Nal-Pro-Arg--k-Gly-Pip	11.6	90.0	
54	D-8-Nal-Pro-Arg--k-Gly-Pip	2.87	5.3	45.0

TABLE 2 - In-vitroassay

	Ki (μM)	TT (μM)*	APTT (μM)	Blood pressure % of normal
	Boc-D-Phe-Pro-Arg-H	0.1	5	40**
	Boc-D, L-Dpa-Pro-Arg-H	0.03	3	100**
	Boc-D, L-Dpa-Pro-Gpa-H	0.03	3	nd
33	Z-D-Phe-Pro-Pgl-H	4.66	86.0	nd
48	Z-D, L-Dpa-Pro-Pgl-H	0.071		nd
53	Boc-D-Phe-Pro-His-H	0.726	3.1	34.0 100***

* The concentration needed to double the plasma thrombin time

** 4mg/Kg given i.v. to anaesthetized cats

*** 1mg/Kg given i.v. bolus to anaesthetized New Zealand white rabbits, 2 per point

ND = Not determined.

TABLE 3

	Ki (uM)
Boc-D-Phe-Pro--ArgCN	0.8
Boc-D, L-Dpa --"	0.05

SUBSTITUTE SHEET

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TABLE 4 - In-vitroassay

	Ki(μM)	TT(μM)
15 Boc-D, L-Dpa-Pro-Pgl ^P (OH) ₂	59.0	
16 D, L-Dpa-Pro-Pgl ^P (OH) ₂	0.223	
17 Z-D-Dpa-Pro-Pgl ^P (OPh) ₂	8.46	
18 D-Dpa-Pro-Pgl ^P (OH) ₂	15.1	
19 Z-D-Phe-Pro-Pgl ^P (OPh) ₂	>31.0	
20 D-Phe-Pro-Pgl ^P (OPh) ₂	0.109	
21 D-Phe-Pro-Pgl ^P (OH) ₂	>98.0	
23 D-Dpa-Pro-Pgl ^P (OPh) ₂	0.48	
32 H-D-Dpa-Pro-Mpg ^P (OPh) ₂	0.0048	9.1
40 H-D-Phe-Pro-Mpg ^P (OPh) ₂	0.0188	

TABLE 5 - In-vitroassays

	Ki(pM)*	TT(μM)	APTT(μM)
12 Z-D-Dpa-Pro-BoroIrg-OPin	368.0	0.84	1.19
26 Z-D-β-Nal-Pro-BoroIrg-OPin	325.0	0.15	0.216
36 Z-D-Fgl-Pro-BoroIrg-OPin	1750.0	0.643	1.23
37 Ac-D-Dpa-Pro-BoroIrg-OPin	2320.0		0.156
38 Z-L-Dpa-Pro-BoroIrg-OPin	4180.0	0.26	0.239
46 Z-D-Cha-Pro-BoroIrg-OPin	1490.0	0.113	0.123

*Inhibitor Pre-Incubated for 30 min with Thrombin.

TABLE 6 - In-vitroassays

	Ki(nM)	TT(μM)	APTT(μM)
22 Z-D-Phe-Pro-BoroMbg-OPin	7.0	0.56	4.11
41 Z-D-Phe-Pro-BoroPhe-OPinac	12.8	0.329	2.52
10 Z-D-Phe-Pro-BoroAcet-OPinac	25.0	3.4	18.2
11 Z-D-Phe-Pro-BoroPgl-OPinac	19.0	2.0	2.34
43 Z-D-Phe-Pro-BoroOct-OPinac	22.5	3.86	
51 Z-D-Dpa-Pro-BoroMpg-OPin	3.4	0.629	1.96
59 Z-D-Phe-Pro-BoroMbg-OPin	7.43	1.12	4.0

TABLE 7 - In-vitroassay

	Km(μM)
H-D-Phe-Pip-Arg-pNA	10.0
34 H-D-β-Nal-Pip-Arg-pNA	6.9
35 H-D, L-Dpa-Pip-Arg-pNA	9.1

SUBSTITUTE SHEET

Those compounds of the invention which are thrombin inhibitors have anti-thrombogenic properties and may be employed for indications when an anti-thrombogenic agent is indicated. Generally, these compounds may be administered orally or parenterally to a host to obtain an anti-thrombogenic effect. In the case of larger mammals such as humans, the compounds may be administered alone or in combination with pharmaceutical carrier or diluent at a dose of from 0.02 to 15 mg/Kg of body weight and preferably 1-10 mg/Kg to obtain the anti-thrombogenic effect, and may be given as single dose or in divided doses or as a sustained release formulation. When an extracorporeal blood loop is to be established for a patient, 0.1-1mg/Kg may be administered intravenously. For use with whole blood from 1-10mg per litre may be provided to prevent coagulation. Pharmaceutical diluents are well known and include sugars, starches and water which may be used to make tablets, capsules, injectable solutions and the like. The compounds of the invention may be added to blood for the purpose of preventing coagulation of the blood in blood collecting or distribution containers, tubing or implantable apparatus which comes in contact with blood.

The advantages of the compounds of the invention include oral activity, rapid onset of activity and low toxicity. In addition, these compounds may have special utility in the treatment of individuals who are hypersensitive to compounds such as heparin.

In the following examples, the symbols have the following meanings:

Aa	=	amino acid
Ac	=	acetyl
Boc	=	t-butyloxycarbonyl
Bu	=	butyl
Bzl	=	benzyl
DGC	=	dicyclohexylcarbodiimide
DIEA	=	diisopropylethylamine

DMAP	=	4-dimethylaminopyridine
EtOAc	=	ethyl acetate
EtOH	=	ethylalcohol
HOSu	=	N-hydroxy succinimide
MCA	=	4-methyl-coumaryl-7-amide
MeOH	=	methylalcohol
Mtr	=	4-methoxy-2,3,6-trimethylbenzenesulphonyl
NMR	=	nuclear magnetic resonance
NP	=	p-nitrophenyl
PinOH	=	pinanediol
PfpOH	=	pentafluorophenol
pip	=	piperidide
pNA	=	p-nitroanilide
TLC	=	thin layer chromatography
THF	=	tetrahydrofuran
TEA	=	triethylamine
WSC	=	water soluble carbodiimide
Z	=	Cbz=benzyloxycarbonyl
Apa	=	amidinophenylalanine
Chg	=	cyclohexylglycine
Dpa	=	3,3-diphenylalanine
Gpa	=	guanidinophenylalanine
Irg	=	isothiuronium analogue of Arg
ArgCN	=	Arg, where COOH is replaced by CN
Mbg	=	2-(2-methylbutyl) glycine
Nal	=	naphthylalanine
Pgl	=	pentylglycine
Thi	=	thiazolidinecarboxylic acid

boroAa	=	boronic acid analogue of Aa
Aa ^P	=	phosphonic acid analogue of Aa
-k-	=	amide bond replaced by CO-CH ₂

The following non-limiting examples illustrate the preparation of the compounds in this invention.

The synthesis of some of the different inhibitor types are outlined in Schemes 1 to 8 and the detailed descriptions are given in the examples below.

HPLC

The following conditions were adopted for the analysis of most of the synthetic compounds on reversed-phase HPLC (RP-HPLC): column; SuperPac Pep-S (4x250mm), eluant; A = water containing 0.1% TFA, B = acetonitrile containing 0.1% TFA, gradient; 50% to 90% B in A in 25 min, flow rate; 1.0ml/min, detection; UV absorbance at 210 nm.

TLC

Thin layer chromatography (TLC) was carried out on the following compounds using precoated silica plates (Merck, F254) in the following systems: A, Chloroform-ethyl acetate (2:1); B, chloroform-methanol-acetic acid (20:4:1); C, n-butanol-acetic acid-ethyl acetate-water (1:1:1:1); D, chloroform-methanol (9:1); E, pyridin-ethyl acetate-acetic acid-water (5:5:1:3); F, chloroform-methanol-ammonia (1M) (60:35:5). The spots were visualized by ninhydrin and chlorine-dicarboxidine spray reagents (C.M. Swahn and J. Gyllander, J. Chromatogr. (1979) 170, 292:

NMR spectra

Magnetic resonance spectra were recorded at 250 MHz using a Bruker instrument.

EXAMPLES1. Synthesis of Dpa, Z-Boc-Dpa-Pro and Z-Dpa-Pro-Arg (Cf. Scheme 1)(a) DL-Dpa.HCl

To a solution of potassium tertiary butoxide (6.75g, 0.06 mol) in tertiary butanol (350ml) was added, at room temperature under argon, ethyl acetamido cyanoacetate (10g, 0.059 mol). When the solution had become clear bromodiphenyl methane (14.55g, 0.059 mol) was added. The mixture was stirred at 20°C for 24h, then evaporated under reduced pressure. The solid residue was treated with ethyl acetate (500ml) and water (175ml). The organic phase was dried (Na_2SO_4) and concentrated to give yellow crystals. The crystals were washed repeatedly with ether and dried to give ethyl 2-diphenylmethylacetamido cyanoacetate (11.61g, 58%, m.p. 181-185°C). The ester (11.61g, 34.4 mol) was mixed with hydrochloric acid (20%) and refluxed for 30h. The reaction mixture was allowed to cool and the crystals were collected, washed (ether), and dried to give HCl.D,L-Dpa (7.82g, 81.8%).

(b) Z-DL-Dpa

To a solution of D,L-Dpa.HCl (0.56g, 0.0021 mol) in NaOH (2N, 5ml), cooled to 0°C and vigorously stirred was added, dropwise, benzyl chloroformate (0.39g, 0.33ml, 0.0023mol). The reaction mixture was kept at pH10 and at 5°C to 10°C. The solution was warmed to room temperature and stirred vigorously for 1h. The solution was washed with ether (4 times) and acidified to pH3 with HCl (5N). The mixture was extracted with dichloromethane and the organic phase dried and concentrated to give Z-DL-Dpa (0.73g, 97%, m.p. 214-217°C).

(c) Z-D,L-Dpa-ONSu

To a stirred solution of Z-D,L-Dpa (1.88g, 0.005 mol) and N-hydroxy succinimide (0.575g, 0.005 mol) in dry 1,2-dimethoxyethane (30ml) at 0°C was added dicyclohexyl carbodiimide (1.03g, 0.005 mol). The mixture was maintained at 0°C for 4h. The suspension was filtered and the filtrate was concentrated to dryness to give an oil which was triturated with ether and filtered to give Z-D,L-Dpa-ONSu (2.15g, 91%, m.p. 139-142°C).

(d) Z-D-Dpa-Pro and Z-L-Dpa-Pro

To a solution of proline (0.78g, 0.0068mol) and NaHCO_3 (0.57g, 0.0068mol) in water (8ml) was added a solution of Z-D,L-Dpa-ONSu (2.15g, 0.0045mol) in 1,2-dimethoxyethane (15ml). After 2h the solvent was removed under reduced pressure and water (5ml) was added. The solution was acidified (conc. HCl) to pH2 to give white crystals (1.98g, m.p. 113-117°C). Fractional recrystallisation from EtOAc gave as the first crop one diastereomer as a solid (0.7g, m.p. 180-183°C, FAB MS: M^+ 473; ^1H nmr: 7.26 (15H, m, 3xPh), 5.66 (1H, d, CH), 5.23 (1H, m, CH), 4.40 (1H, d, CH), 2.03 (2H, s, CH_2), 2.20 (4H, m, 2x CH_2); ^{13}C nmr: 172.19 (CO), 156.1 (CO), 139.17 (CO), 127-128 (Ph), 66.88 (CH_2), 59.48 (CH), 55.58 (CH), 24.15 (CH_2). Further crystallisation from the mother liquor gave as a second crop a mixture of diastereomers (0.43g, m.p. 126-130°C). Addition of petroleum ether (b.p. 60-80°C) gave the other isomer (0.54g, m.p. 128-131°C), FAB MS: M^+ 473; ^1H nmr: 7.29 (15H, m, 3Ph), 5.55 (1H, d, CH), 5.23 (1H, m, CH), 4.47 (1H, d, CH), 2.04 (2H, s, CH_2), 1.20-2.20 (4H, m, 2 CH_2); ^{13}C : 172.77 (CO), 156.13 (CO), 139.48 (CO), 126.99-128.72 (Ph), 66.90 (CH_2), 59.62 (CH), 55.48 (CH), 53.54 (CH), 47.44 (CH_2), 27.94 (CH_2), 24.58 (CH_2).

(e) Z-D-Dpa-Pro-Arg(Mtr)OPh

To a solution of Z-D-Dpa-Pro-OH (0.472g, 1mmol) and HOSu (0.115g, 1mmol) in dimethoxyethane (20ml) was added DCC (0.206g, 1mmol) with cooling over an ice water bath, the solution was then stirred at r.t. for 3h, the DCU formed was filtered off and the solution was concentrated to dryness to give an oil (0.57g). To a solution of H-Arg(Mtr)-OH (0.42g, 1.1mmol) and Et_3N (0.12g, 1.1mmol) in DMF (25ml) was added a solution of Z-Dpa-Pro-OSu (0.57g) in dimethoxyethane (15ml) with cooling. The solution was stirred at room temperature for 3h. The solvent was evaporated and the residue was dissolved in H_2O (20ml) and MeOH (10ml). The solution was acidified to pH2 and the MeOH was removed under reduced pressure. The solid formed was filtered off and

dried to give Z-D-Dpa-Pro-Arg(Mtr)OH (0.766g, 91%). The structure of the compound was confirmed by ^1H NMR.

Pgl and Nal and their corresponding di- and tripeptides were synthesized in a manner analogous to the above procedures.

2. Synthesis of peptide aminophosphonic acid inhibitors (Cf. Scheme 3)

(a) Diphenyl 1-(N-benzyloxycarbonyl)aminopentanephosphonate

A mixture of triphenyl phosphite (9.3g, 30mmol), n-hexanal (4.50g, 45mmol), benzylcarbamate (4.53g, 30mmol), glacial acetic acid (5ml) was stirred for 45min. The mixture was then heated at 80-85°C for 1h and volatile by-products were removed in vacuo with heating on a boiling water bath. The oily residue was dissolved in methanol (40ml) and left for crystallization at -10°C to give 7.28g, m.p. 70-72°C, 52% yield. The structure was confirmed by proton NMR.

(b) Diphenyl 1-aminopentanephosphonate

Diphenyl-1-(N-benzyloxycarbonyl) aminopentanephosphonate (0.93g, 2.0mmol) was dissolved in ethanol (30ml) and acetic acid (0.2ml) was added. Then 10% palladium on charcoal (100mg) was added and the mixture was hydrogenated for 4h. The catalyst was filtered off, washed with ethanol (5x5ml). After removal of the solvent an oil was obtained. The oil was washed with water to remove acetic acid and dissolved in chloroform, dried (MgSO_4), concentrated to dryness to give oily product, 0.45g, 68% yield. The structure was confirmed by proton NMR and MS.

(c) Z-D-Dpa-Pro-PglP (OPh)₂

Z-D-Dpa-Pro-OH (0.11g, 0.25mmol) was dissolved in dry chloroform (2ml) containing Et_3N (0.035ml) and cooled to -5°C. Ethyl chloroformate (0.026ml, 0.275mmol) was added and the mixture kept at -5°C for 30 min. A solution of diphenyl 1-aminopentanephosphonate (83mg, 0.25mmol) in dry chloroform (2ml) containing Et_3N (0.025g, 0.25mmol) was added. The mixture was stirred at r.t. for 12h. Solvent was removed in vacuo. The resulting oil was chromatographed

(CHCl₃ then 2% MeOH in CHCl₃) to give 123mg as crystals, 63% yield. The structure was confirmed by proton and ³¹P NMR.

(d) H-D-Dpa-Pro-Pgl^P (OPh)₂

Z-D-Dpa-Pro-Pgl^P (OPh)₂ (50mg, 0.063 mmol) was dissolved in ethanol (5ml) and acetic acid (0.01ml) was added. 10% Pd/C (25mg) was added and the mixture was hydrogenated at r.t. for 3h. The catalyst was filtered off and ethanol removed in vacuo. The resulting oil was treated with water (5ml) and chloroform (20ml). The chloroform phase was dried (MgSO₄) and concentrated to dryness to give crystals, 41mg, 91% yield. The structure was confirmed by ¹H and ³¹P NMR.

(e) H-D-Dpa-Pro-Pgl^P (OH)₂

Z-D-Dpa-Pro-Pgl^P (OPh)₂ (100mg, 0.127mmol) was dissolved in ethanol (10ml) and acetic acid (0.1ml) was added. Then 10% Pd/C (50mg) was added and the mixture was hydrogenated at r.t. for 3h. The catalyst was filtered off, PtO₂ (100mg) was added and the mixture was hydrogenated at r.t. for 4h. The catalyst was filtered off, solvent was removed and the residue was treated with 20ml of water and chloroform (60ml). The organic layer was dried (MgSO₄), concentrated to dryness to give 67mg as crystals, 92% as overall yield. The structure was confirmed by ¹H and ³¹P NMR.

Z-D-Phe-Pro-Pgl^P (OPh)₂

This compound was synthesized by the above procedure in 73% yield. The structure was confirmed by ¹H and ³¹P NMR.

H-D-Phe-Pro-Pgl^P (OPh)₂

The compound was synthesized by the above procedure in 90% yield. The structure was confirmed by ¹H and ³¹P NMR.

H-D-Phe-Pro-Pgl^P (OH)₂

The compound was synthesized by the above procedure in 89% overall yield. The structure was confirmed by ¹H and ³¹P NMR.

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(f) Diphenyl 1-(N-allyl)amino-4-Pyridylmethyl-phosphonate

To a solution of 4-pyridinecarboxyaldehyde (1.07g, 10mmol) and allylamine (0.61g, 10mmol) in ether (30ml) was added anhydrous sodium carbonate (2.76g). The solution was stirred at r.t. overnight, then sodium carbonate was filtered off. To the reaction mixture, diphenylphosphite (2.34g, 10mmol) and triethylamine (1.01g, 10mmol) were added with cooling over an ice-water bath. It was stirred at r.t. overnight. After removal of the solvent an oily residue was obtained which was chromatographed (1:1 Petroleum ether/ethyl acetate) to give 2.85g (65%) as an oil.

(g) Diphenyl 1-(N-allyl)amino-4-(tert-butylloxycarbonyl)butyl-phosphonate

4-(tert-butylloxycarbonyl)amino-butylaldehyde diethyl acetal (2.91g, 10mmol) was dissolved in acetone (20ml) in the presence of 1N hydrogen chloride (1ml) and PPTS (150mg). The reaction mixture was refluxed for 3h. The solvent was removed and the residue was dissolved in chloroform and dried (MgSO_4). After removal of MgSO_4 the solution was stirred and allylamine (0.61g, 10mmol) and anhydrous sodium carbonate (2.76g) were added. The reaction suspension was stirred at r.t. overnight. Then sodium carbonate was filtered off. To the solution obtained, diphenylphosphite (2.34g, 10mmol) and the triethylamine (1.01g, 10mmol) were added. It was stirred at r.t. for 2 days. The residue obtained after evaporation was chromatographed on silica gel (1:1 Petroleum/ethyl acetate) to give 200mg as a yellow waxy

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solid (5%).

(h) Diphenyl 1-amino-4-pyridyl-methyl-phosphonate

The N-allyl protected compound (1.0g, 2.3mmol) was dissolved in ethanol (25ml). To the solution was added 10% Pd/C (300mg) and it was refluxed for 20 hours. The reaction was followed by HPLC, retention time: 10.0min for the product and 12.0min for the starting material. After removal of the solvent the product was obtained by chromatography, 0.51g (56%) as a yellow oil.

The following tripeptides were synthesized according to Scheme 5:

Z-D-Phe-Pro-Cpg^P(OPh)₂: 0.36g (67%)
Z-D-Phe-Pro-Epg^P(OPh)₂: 0.31g (87%)
Z-D-Phe-Pro-Pyg^P(OPh)₂: 0.41g (35%)
Z-D-Phe-Pro-Dmg^P(OPh)₂: 0.25g (70%)
Z-D-β-Nal-Pro-Mpg^P(OPh)₂: 54mg (32%)
H-D-Phe-Pro-Epg^P(OPh)₂: 126mg (71%)
H-D-Phe-Pro-Dmg^P(OPh)₂: 85mg (52%).

Abbreviations:

Mpg = methoxypropylglycine
Cpg = 4-cyanophenylglycine
Epg = 2-ethylpropylglycine
Pyg = 4-pyridylglycine
Dmg = 3,3-dimethylpropylglycine
Nal = naphthylalanine
PPTS = Pyridinium p-toluenesulfonate

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3. Synthesis of peptide aminoboronic acid inhibitors

(a) (+)-Pinanediol 4-bromo-R-1-aminobutane boronate hydrochloride

The title compound was prepared as described by D.S. Matteson et al (1984) in *Organometallics*, **3**, 1284-1288 and in European patent appl. 293881A2.

(b) Z-D-Dpa-Pro-Irg-OPin.HCl

To a solution of Z-D-Dpa-Pro-OH (236mg, 0.5mmol) in THF (5ml) in the presence of triethylamine (70µl, 0.5mmol) was added isobutyl chloroformate (65µl, 0.5mmol) at -15°C and the solution was stirred at -13°C for 13 min. After the addition of (+) - pinanediol 4-bromo-R-1-aminobutaneboronate hydrochloride (183mg, 0.5mmol) in CHCl₃ (3ml) followed by that of Et₃N (70µl, 0.5mmol), the reaction mixture was stirred at the same temperature for 2h, and then below 10°C for 2h. THF was removed under reduced pressure and the residue was dissolved in ethyl acetate (50ml), which was washed with 1% NaHCO₃, water, 0.2N HCl and water, and then dried over Na₂SO₄. Removal of solvent gave an oily product quantitatively. HPLC analysis showed one major peak at the retention time of 22.8 min along with several minor components.

To a solution of the above compound (2/5th of total amount synthesized, 0.2mmol) in ethanol (1ml) was added thiourea (61mg, 0.8mmol) under an atmosphere of argon at room temperature. After stirring for 4 days, ethyl acetate (70ml) was added to the reaction mixture, which was washed with 1% NaHCO₃, water, 0.2N HCl and then water, and dried over Na₂SO₄. The residue obtained by removing the solvent was treated with n-hexane to get the product as a powder. Reprecipitation from ethyl acetate with 2:1 mixture of ethyl ether and n-hexane gave a product (98.9mg, 60.6%, two step overall). Retention time on RP-HPLC analysis was 13.5 min under the conditions described at the general procedure. ¹H NMR analysis in deuterated chloroform gave a complex pattern because of the existence of proline residue in the molecule, however, the typical signals corresponding to pinanediol were observed as proper ratios.

4. Z-D-Phe-Pro-boromBg-OPin (Cf. Scheme 4)

A solution of pinanediol (dichloromethyl)boronate (1ml, 1.2g, 4.6mmol) in THF (7ml) was placed in a septum fitted flask (100ml), and 1,1-dimethylpropane magnesium chloride (4.6ml, 4.6mmol) added dropwise from a dry syringe at 0°C.

The reaction mixture was left stirring under nitrogen at room temperature. After 7 hours TLC showed mainly one spot [$R_f = 0.82$, chloroform:pet.ether (1:1)]. The solvent was removed and the residue dissolved in ether (50ml), washed with water (2x10ml), dried ($MgSO_4$) and filtered.

The ether was removed and the crude product purified on a column of silica gel, eluted with hexane and 10% of chloroform to give the α -chloroboronic ester as a pale yellow oil (0.55g, 40% yield).

The above compound (0.55g, 1.8mmol) in THF (5ml) was added via a double ended needle at -78°C to a solution of lithiumbis(trimethyl-silyl) amide (1.8ml, 1.8mmol) in THF (5ml) under nitrogen. The reaction mixture was kept overnight at 20°C then the solvent was removed. The crude product was dissolved in petroleum ether (40-60°C) (25ml) to precipitate out the inorganic salt ($LiCl$). The reaction mixture was filtered, cooled to -78°C and dry ethereal HCl , 1M (3 equiv, 5.4ml, 5.4mmol) added. The flask was kept in a fridge overnight. Next morning, the reaction mixture was filtered to isolate the hydrochloride (0.41g, 1.29mmol, 72% yield) as a white solid.

Z-D-Phe-Pro-OH (0.45g, 1.1mmol) was dissolved in THF (7ml) and the equivalent of N-methylmorpholine (0.11g, 1.1mmol) added. The solution was cooled to -20°C and one equivalent of isobutylchloroformate (0.149g, 1.1mmol) added dropwise. After 10 min., a solution of the above aminohydrochloride (0.348g, 1.1mmol) dissolved in THF (7ml) was transferred under nitrogen, and triethylamine (0.11g, 1.1mmol) added to the reaction mixture. The reaction mixture was stirred for one hour at -20°C, followed by 2h at room temperature. Insoluble material was removed by filtration, then the solvent removed by

evaporation, and the residue dissolved in ethyl acetate (30ml). The organic layer was washed with 0.2N hydrochloric acid (10ml), 5% aqueous sodium bicarbonate, saturated solution of sodium chloride and water. The organic phase was then dried over anhydrous MgSO_4 , filtered and the solvent evaporated to give a white solid which was purified on a column of silica gel eluted with light petroleum to give the desired product (0.59g, 81%). The structure was confirmed by ^1H NMR and MS.

5. PREPARATION OF α -BROMO BORONIC ESTERS (Cf. Scheme 6)

All the reactions involving boron used purified anhydrous reagents.

Reactions were carried out under argon or nitrogen used directly from the cylinder through a glass line.

In a 250 ml reaction flask fitted with a reflux condenser was placed 1-bromo-1-propene (3.63g, 30mmol).

Dibromo borane -methyl sulfide complex in dichloromethane (60ml, 60mmol) was then added to the reaction flask dropwise and the mixture was refluxed under nitrogen for 5h. The solvent was removed and the reaction mixture washed with water and dried (MgSO_4).

A dry round-bottomed flask (100ml) was charged with the bromo boronic acid (0.5g, 3mmol) and pinanediol (0.52g, 3mmol) a magnetic follower and dry ether (20ml), fitted with a septum and flashed with nitrogen.

The reaction mixture was left stirring for two hours until the solid dissolved, the organic phase was washed with water (10ml), separated, dried (MgSO_4) and filtered. The crude product was purified on a column of silica gel (230-400 mesh), eluted with chloroform (the product was eluted before the pale red ring). The first fraction (100ml) was collected and the solvent evaporated to give α -bromo boronic ester (0.8g, 88.6%) as a colourless liquid.

6. Synthesis of isosteric ketomethylene inhibitors (Cf. Scheme 2)

(a) Boc-D-Dpa-Pro-Arg(Mtr)-k-GlyOMe (modified Darkin-West reaction)

Boc-D-Dpa-Pro-Arg(Mtr)-OH (0.126g, 0.15mmol) was added to monomethylsuccinyl anhydride (0.259g, 1.0mmol). Et_3N (0.042ml, 0.30mmol), DMAP (1.8mg, 0.015mmol) and pyridine (0.12ml) were added, the reaction flask was fitted with a reflux condenser, and the reaction was heated at 45-50°C. The reaction mixture was stirred for 1h, then NaHCO_3 (5%, 5ml) was added and the stirring was continued for an additional 30min. The product was extracted into ethyl acetate and washed with AcOH (0.1N) and brine. The organic layer was dried (MgSO_4), filtered and concentrated to dryness to give an oily residue which was chromatographed on silica gel (grade 9385, 50g). Elution with $\text{CHCl}_3:\text{CH}_3\text{OH}$, 98:2 gave the product after removal of the solvent as a brown oil (0.134g, 98%). The structure was confirmed as Boc-Dpa-Pro-Arg(Mtr)-k-GlyOMe by ^1H NMR (250MHz), and by FAB mass spectrometry.

(b) Z-D-Dpa-Pro-Arg(Mtr)-k-Gly-pip

A solution of Z-D-Dpa-Pro-Arg(Mtr)-k-Gly-OMe (0.1g, 0.1mmol) in MeOH (10ml) was cooled to 0°C and NaOH (1N, 0.22ml, 0.22mmol) was added with stirring for 2.5h at room temperature. The solution was neutralized to pH7 and the MeOH was removed under reduced pressure. The aqueous solution was acidified (pH2) and extracted by ethyl acetate and dried (Na_2SO_4). The solvent was removed under reduced pressure to give an oil. To a solution of the oil and HOSu (12mg, 0.1mmol) in dimethoxy ethane (20ml) was added DCC (21mg, 0.1mmol), with

cooling. The solution was stirred at room temperature for 20h, and piperidine (17mg, 0.2mmol) was added to the solution with cooling and the solution was stirred at room temperature for a further 3h. The solution was concentrated to dryness and the product was purified by chromatography on silica gel (MeOH:CHCl₃, 92:2) to give Z-D-Dpa-Pro-Arg(Mtr)-k-Gly-pip (78mg, 81%). The structure of the product was confirmed by ¹H NMR and FAB mass spectrometry.

In a separate experiment the L-isomer, Z-L-Dpa-Pro-Arg(Mtr)-k-Gly-pip, was synthesized by the above procedure in 55% yield.

(c) H-D-Dpa-Pro-Arg-k-Gly-pip.2TFA

Z-D-Dpa-Pro-ArgMtr-k-Gly-pip (52mg, 0.053mmol) was dissolved in 0.9ml of TFA and 0.1ml of thioanisole at room temperature. After stirring for 4h. TFA was removed at reduced pressure and the residue was triturated with ether. The crystals were collected and washed with ether (51mg of Mtr deprotected product). The crystals were then dissolved in 5ml of methanol and 21mg of 10% Pd/C was added. After 20h. hydrogenation at room temperature the catalyst was removed by filtration and the solvent evaporated. The residue was triturated with ether to give white crystals. 34mg (75%), m.p. 146-151 (dec.). HPLC showed two equally big peaks, from the two forms containing D and L Arg. The structure was confirmed by ¹H NMR and FAB mass spectrometry.

In a separate experiment the L-isomer of Z-Dpa-Pro-Arg(Mtr)-k-Gly-pip was deprotected to give H-L-Dpa-Pro-Arg-k-Gly-pip.2TFA in 43% yield.

7. Synthesis of peptide aldehydes (Cf. Scheme 8)

(a) Z(NO₂)Arg NCH₃(OCH₃)

N,O-dimethylhydroxylamine hydrochloride (1.45g, 14.9mmol) was dissolved in 10ml of DMF, and the solution kept at 0°C. Diisopropylethylamine (1.92g, 14.9mmol) was first added and then Z-(NO₂)Arg in 10ml of DMF, HOBT (1.92g, 14.2mmol) and WSC.HCL (2.99g, 15.6mmol). After 5h reaction at 0°C the solution was left at room temperature overnight. The solvent was removed at reduced

pressure and 100ml of EtOAc and 25ml of H₂O were added. The organic phase was diluted with ether and washed with Na₂CO₃ (0.5M), H₂O, H₂SO₄ (0.1M) and H₂O, then dried and the solvents removed giving 3.66g of product. The water solutions were combined, extracted and treated in the same way as above giving further 1.69g. Totally 5.35g (95%) which was chromatographed on Sephadex LH-20 with 95% ethanol. Yield 4.60g (82%) of homogenous product (TLC in S₁ and S₂). NMR confirmed the structure.

(b) Arg(NO₂)-NCH₃(OCH₃).HBr

The above compound was deprotected in HBr/HOAc at room temperature for 45min in the usual manner. The product was homogenous according to TLC in S₃, S₄ and S₅.

(c) Boc-D,L-Dpa-Pro-Arg(NO₂)-NCH₃(OCH₃)

Arg(NO₂)-NCH₃(OCH₃).HBr (4.9g, 13mmol) was dissolved in DMF, the solution cooled to -5°C and Et₃N added to alkaline reaction. Boc-DL-Dpa-Pro OH (5.5g, 12.5mmol), HOBT (1.7g, 12.5mmol) and WSC (2.8g, 14.5mmol) were added and the reaction mixture stirred for 2h. at -5°C and then stirred at room temperature overnight. The solvent was evaporated at reduced pressure, EtOAc and H₂O were added, the organic phase separated and extracted with 0.5M NaHCO₃ (3x30ml), NaCl solution (4x20ml), dried (NaSO₄) and the solvent removed. Yield 8.3g (97%) TLC (S₂) shows one spot. The expected structure of the compound was confirmed with NMR.

(d) Boc-D,L-Dpa-Pro-Arg-NCH₃(OCH₃).HCl

The above compound (2.33g, 3.4mmol) was dissolved in 240ml of MeOH and 3.6ml HCl (1M). The catalyst, 10% Pd/C (0.6g) was added and hydrogenation performed at room temperature for 20h. The catalyst was filtered and the solvent removed at reduced pressure. Remaining 2.3g solid contained some starting material which was removed by ion exchange chromatography on Sephadex QAE + Cl⁻ with 50% EtOH. Yield 1.74g (76%). TLC (S₂ and S₃) showed a single spot.

(e) Boc-D,L-Dpa-Pro-Arg-H.Cl

The above compound (0.5g, 0.74mmol) was dissolved in 40ml of dried (molecular sieve, 4A) THF and the solution was cooled to -40°C and 3.2ml of DIBAH (1M toluene solution, 3.2mmol) was added dropwise during stirring in argon atmosphere. After 3h. 12.8ml of 0.25M citric acid was added. The aluminium salts were centrifuged and washed several times with THF/H₂O (4:1). From the combined liquid phases THF was removed at reduced pressure and the product extracted with EtOAc. The solvent was removed, the product dissolved in 15ml of 20% HOAc and chromatographed on Sephadex G15 with 20% HOAc as eluent. Yield 172mg (38%). The compound shows a double spot in TLC (S₂ and S₃) probably showing the two isomeres with D- and L-Dpa, respectively. NMR and MS were in agreement with the expected structure.

8. Synthesis of Boc-D,L-Dpa-Pro-ArgCN.HCl(a) Z-D,L-Dpa-Pro-Arg-NH₂.HCl

Arg-NH₂.2HCl and Boc-D,L-Dpa-ProOH were coupled in the normal way with HOBT and DCC in DMF. Yield 53%.

(b) Boc-D,L-Dpa-Pro-ArgCN.HCl

The above tripeptide amide (0.50g, 0.79mmol) and tosylchloride (0.50g, 2.55mmol) were dissolved in 2ml of pyridine at room temperature and stirred for 24h. The pyridine was evaporated at reduced pressure, 5ml of pyridine and 0.5ml of water were then added and stirring continued for 2h. After evaporation at reduced pressure the residue was triturated with a small amount of water, dissolved in EtOAc, dried (Na₂SO₄) and chromatographed on Sephadex QAE[®]Cl[⊖]. The fractions containing the product were evaporated, dissolved in 10% HOAc and freeze dried. Yield: 0.33g (68%). $[\alpha]_{\text{D}}^{22} = -144^{\circ}$ (C=0.5, 50% HOAc). The structure was confirmed with ¹H NMR and mass spectroscopy.

9. Synthesis of Db

(a) Preparation of N-formyl-N, α , β -tribenzylalanine cyclohexylamide

Bzl-NH₂ 288 μ l (2.64 mmol) and benzyl phenethylketone 592 mg (2.64 mmol) were dissolved in MeOH (5 ml) at rt and the solution was stirred overnight. To the mixture were added formic acid 99.6 μ l (2.64 mmol) and cyclohexylisocyanide 298 μ l (2.4 mmol) at rt, which was allowed to react at rt for 2 weeks. Insoluble material in MeOH was collected by filtration and washed with MeOH, ether and then n-hexane. The crude product was recrystallized from CHCl₃ with the 2 to 1 mixture of ether and n-hexane. Yield 540 mg (48.2%), NMR in CDCl₃; δ =0.8-1.8 cyclohexyl (11H), δ =2.1-4.75 CH₂ (8H), δ =5.45-5.55 NH (1H), δ =7.0-7.4 phenyl (15H), δ =8.25 (main) and 8.4 (minor) formyl (1H).

(b) Preparation of Bzl-Db

The fully protected Db 400 mg (0.85 mmol) was dissolved in 2.5 ml of TFA and 3 ml of 11 N HCl and the solution was kept at 145°C with water-cooled condenser and stirred for 20 h. After removal of TFA, the pH of the solution was adjusted around 7 by addition of 10 N NaOH. Further addition of ether gave a powder which was collected and washed with ether. Yield 124 mg (40.4%).

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10. Synthesis of Z-D-Phe-Pro-Pgl-H(a) Pgl

Pentylglycine was obtained by the Strecker* synthesis, using hexanal, in a yield of 31.6% (*Vogel, Textbook of Practical Organic Chemistry).

(b) Z-Pgl

To a solution of pentylglycine (0.5g, 3.5mmol) in a mixture of water (4ml) and THF (4ml) to give a 0.5M solution, in the presence of triethylamine (0.64ml, 1.22eq.) was added Z-OSu (0.963g, 3.86mmol.) at room temperature, solution became clear after 15min. TLC after 2h indicated some starting material and so a further portion of triethylamine (0.2ml) and Z-OSu (200mg) was added. TLC after a further 2h indicated no starting material and so the solution was poured onto water (50ml) and was extracted by CHCl_3 (50ml). The organic phase was washed by HCl (1M, 20ml) and dried (MgSO_4) and concentrated to give Z-Pgl (1.18g) as a gummy solid, which was recrystallised (DCM/Petroleum ether bp60-80°C) to a white crystalline solid (0.8g). Structure was confirmed by ^1H Nmr.

(c) Z-Pgl-NMe(OMe)

To a solution of hydroxylamine (184mg, 1.05eq.) was added DIPEA (0.33ml, 1.05eq.), and after 5min a solution of Z-Pgl (0.5g, 1.8mmol) then HOBT (0.242mg, 1 eq.). The solution was cooled to -15°C and a solution of WSCI.HCl (0.378mg, 1.1eq.) was added. The solution was maintained at -15°C for 30min. then allowed to warm to room temperature and pH adjusted to ~ pH4 by addition of acetic acid. TLC after 16h shows little

starting material and so the solution was poured onto NaHCO_3 (100ml) and extracted by Et_2O (50ml). The Et_2O phase was washed by NaCl (20ml). The aqueous phase was washed by Et_2O , the organic phases were combined and then concentrated to give Z-PglNMe(OMe) (324mg). Structure confirmed by ^1H Nmr and mass spec.

(d) PglNMe(OMe)

To a solution of Z-PglNMe(OMe) (324mg) in MeOH (10ml) was applied a vacuum, then argon. Purging was repeated twice, but on the third time Pd/C (~0.5g) was added. Purging was repeated twice more and on the third time the vacuum was quenched by bubbling Hydrogen through the solution. AcOH (0.5ml) was then added. After 90min TLC indicated no starting material and so the solution was filtered (celite), washing with a large volume of MeOH, and concentrated to give PglNMe(OMe)(380mg), as a gum. Structure was confirmed by ^1H Nmr.

(e) Z-D-PheProPglNMe(OMe)

To a solution of Z-D-PhePro (0.187mg, 1eq.) in DMF (2ml) was added DIPEA (0.084ml, 1eq.) and PyBOP (0.25mg, 1eq.), with stirring under argon. After 10min a solution of PglNMe(OMe) (0.09mg, 0.479mmol), in DMF (1ml), was added. After 90min TLC indicated no starting material and so the solution was poured onto HCl (1N, 50ml) and extracted by Et_2O (50ml). The Et_2O layer was washed by NaHCO_3 (50ml, 1.2N) and NaCl (sat'd, 20ml) and dried (MgSO_4). Repeated chromatography on silica gel (Merck 9385), eluting with $\text{CHCl}_3/\text{MeOH}$ gave Z-D-Phe-Pro-PglNMe(OMe) (200mg). Fab Ms shows 567 (10%, M+H),

and 589 (100%, M+Na) as required, the structure was also confirmed by ^1H Nmr.

(f) Z-D-Phe-Pro-Pgl-H

To a solution of Z-D-PheProPglNMe(OMe) (33mg) in THF (2ml) at - 40°C was added Di-isobutylaluminium hydride (1N, 0.155ml, 2.5eq.), under argon. The solution was allowed to warm to room temperature and stirred for 18h. TLC showed no starting material and so was quenched by H_2SO_4 (1N, 0.5ml) and stirred for 10min. The aqueous phase was then extracted by EtOAc (20ml). the organic phase was dried (MgSO_4) and concentrated to give Z-D-PheProPgl-H (31mg). The structure was confirmed by ^1H Nmr and mass spectrum, and the compound was sent for biological testing, as compound number 33.

11. Z-L-Val-pNa

4-nitroaniline (67.5g, 0.48mol) was dissolved in pyridine (dried over 4A sieves, 750ml), and the solution was cooled down with ice. PCl_3 (34.3g, 0.25mol) was dissolved in pyridine (350ml) and added dropwise to the nitroaniline solution. The solution was allowed to stand for 30 min. at room temperature. To the solution was added a solution of ZZVal-OH (112g, 0.44mol) in pyridine (250ml). The reaction mixture was stirred at room temperature for 1 week, then the pyridine was removed on a rotavapor. The residue was treated with NaHCO_3 (2%). The product crystallized and was filtered and washed with water. The product was dissolved in boiling EtOH (2000ml, 95%) and the hot solution was filtered and left to stand overnight at room temperature. The crystals were filtered giving Z-L-Val-pNA (116.9g, 71.6%).

12. (a) Boc-Dpa

Boc-Dpa was obtained in an analogous manner to the procedure of Examples 1(a) and (b).

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(b) Boc-Dpa-Pip-Arg-pNA (Cf. Scheme 7)

To a solution of Boc-Dpa.HCl (150mg, 0.396mmol) in DMF(5ml) was added TBTU (133.5mg, 0.416mmol, 1.05eq.) and DIPEA (0.069ml, 0.396mmol, 1eq.), to give a solution of basic pH. PipArgpNA.TFA (238mg, 0.396mmol, 1eq.), was then added. pH of the solution was acidic so a further portion of DIPEA (0.05ml) was added. After stirring for 16h, TLC showed a little starting material and so a further portion of DIPEA was added (0.034ml, 0.5eq.). TLC after a further 3h showed no starting material and so the solution was poured onto EtOAc (50ml) and washed by HCl(0.5N, 200ml), dried (MgSO₄) and concentrated to give 430mg of gum. Trituration with Et₂ and petroleum ether (b.p.60-80°C), over 5 minutes, gave a powder which was filtered, avoiding drying in the air, to give a BocDpaPipArgpNA.TFA as a powder, 207mg, 62% yield, R_f 20min on Rp HPLC (pep-S, 35-70% MeCN+0.1%TFA, 25min, 1ml/min.).

(c) Dpa-Pip-Arg-pNA

To solid BocDpaPipArgpNA (100mg, 0.119mmol), was added TFA (2ml), with cooling in an ice bath for 10min. The ice bath was then removed and the solution stirred for a further 10 min. TLC indicated no starting material, so the solution was concentrated under vacuum (oil pump). Washing with Et₂O(150ml), until the filtrate was neutral pH, gave a powder dried under vacuum to DpaPipArgpNA.2TFA, 92mg, (90%), as a powder, of retention time (15.5min on Rp HPLC pep-s, 4x250mm, 35-70% MeCN +0.1% TFA).

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(d) Boc-D-Nal-Pip-Arg-pNA

To a solution of Boc- -D-Nal(150mg, 0.474mmol) in DMF (5ml) was added TBTU (164mg, 0.51mmol, 1.05eq.) and DIPEA (0.083ml, 0.474mmol, 1 eq.). PipArgpNA.TFA (285mg, 0.474mmol, 1 eq.) was then added. After 30 min TLC, using the Sakaguchi reagent(8-hydroxyquinoline, Br₂, NaOH), indicated mainly starting material, and so DIPEA (0.083ml, 1eq.) was added. After 30min. TLC indicated mainly starting material and so the reaction was diluted by EtOAc (50ml) and washed by HCl (0.5N, 100ml). The organic phase was dried (MgSO₄), and concentrated to give an oil (490mg). Recrystallisation from CHCl₃/petroleum ether bp 60-80°C/Et₂O gave 90mg, 20.1% yield, of Boc-Dpa-PipArgpNA.TFA as a powder.

(e) H-D-NalPipArgpNA.2TFA

To solid Boc- -D-NalPipArgpNA.TFA (50mg, 0.055mmol), cooled in an ice bath, was added TFA (2ml), with stirring. After 10min the ice bath was removed, TLC after 30 min still showed some starting material and so solution was stirred for a further 10 min, then concentrated (oil pump). Trituration with Et₂O gave a powder. The powder was washed with Et₂O, until the eluant was neutral, dried overnight at room temperature to give H-D-NalPipArgpNA as a powder, 42mg, 92% yield. Retention time 11.5min on Rp HPLC.

Example 13

The following compounds were prepared by the route substantially as outlined in Examples 2a to 2e:

	Yield	Physical data to confirm structure
15 Boc-D,L-Dpa-Pro-Pgl ^P (OH) ₂	96%	Nmr, Ms.
16 D,L-Dpa-Pro-Pgl ^P (OH) ₂		
17 Z-D-Dpa-Pro-Pgl ^P (OPh) ₂		
18 D-Dpa-Pro-Pgl ^P (OH) ₂	92%	Nmr, FABMs[M+H] 502, m.p. 130-133.
19 Z-D-Phe-Pro-Pgl ^P (OPh) ₂		
20 D-Phe-Pro-Pgl ^P (OPh) ₂	35%	Nmr, FABMs.
21 D-Phe-Pro-Pgl ^P (OH) ₂		
23 D-Dpa-Pro-Pgl ^P (OPh) ₂		
32 H-D-Dpa-Pro-Mpg ^P (OPh) ₂		H, ³¹ PNmr, HPLC.
40 H-D-Phe-Pro-Mpg ^P (OPh) ₂		HPLC.
55 H-D-Phe-Pro-Apg ^P (OPh) ₂		HPLC.
56 H-D-Phe-Pro-Epg ^P (OPh) ₂	71%	
57 H-D-Phe-Pro-DPg ^P (OPh) ₂	52%	Nmr.

Example 14

The following compounds were synthesised according substantially to Examples 2a to 2d:

Z-D-Phe-Pro-Epg ^P (OPh) ₂	87%
Z-D-Phe-Pro-Cpg ^P (OPh) ₂	67%
Z-D-Phe-Pro-Pyg ^P (OPh) ₂	35%
Z-D-β-Nal-Pro-Mpg ^P (OPh) ₂	32%

Example 15

The following compound was synthesised according to Examples 2a and 2b:
MTy^P(OPh)₂

Example 16

The following compounds were synthesised according substantially to Example 2f:

D-Phe-Pro-Pyg ^P (OPh) ₂
D-Phe-Pro-Aeg ^P (Boc)(OPh) ₂
D-Phe-Pro-Npg ^P (OPh) ₂

Example 17

The following compounds were synthesised according substantially to Examples 6a to 6c:

4 D,L-Dpa-Pro-Arg--k--Gly-Pip		
5 D-Phe-Pro-Gpa--k--Gly-Pip	81.2%	Nmr, FABMs, m.p. 110-114.
6 D-Dpa-Pro-Arg--k--Gly-Pip	74.5%	Nmr, FABMs, m.p. 146-151.
7 L-Dpa-Pro-Arg--k--Gly-Pip	42.7%	Nmr, FABMs, m.p. 136-140.
8 D-Fgl-Pro-Arg--k--Gly-Pip	81.3%	Nmr, FABMs.
9 D,L-α-Nal-Pro-Arg--k--Gly-Pip	81%	Nmr, FABMs, m.p. 123-127.
14 D,L-β-Nal-Pro-Arg--k--Gly-Pip	55%	FABMs.

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54 D-8-Nal-Pro-Arg--k--Gly-Pip 41% FABMs.

Example 18

The following compounds were synthesised according substantially to Example 10:

33	Z-D-Phe-Pro-Pgl-H	99.0%	Nmr, FabMs[M+H]508,7%
48	Z-D-Dpa-Pro-Pgl-H	quantitative	Nmr, FabMs[M+H]584,35%
53	Boc-D-Phe-Pro-His-H	90%	Nmr, FabMs[M+H]484
	Z-N-Me-Phe-Pro-Pgl-NMe(OMe)		

Example 19

The following compounds were synthesised according substantially to Example 11:

34	H-D-8-Nal-Pip-Arg-pNA	90.6%	Nmr
35	H-D,L-Dpa-Pip-Arg-pNA	81%	Nmr
	H-D-Phe-Pro-Phe-pNA		

Example 20

The following compounds were synthesised according substantially to Example 4:

22	Z-D-Phe-Pro-BoroMbg-OPin	89%	Nmr
41	Z-D-Phe-Pro-BoroPhe-OPinac	68.6%	Nmr
59	Z-D-Phe-Pro-BoroMbg-OPin	90%	Nmr

Example 21

The following compounds were synthesised according substantially to Example 3a:

10	Z-D-Phe-Pro-BoroAcet-OPinac	42%	Nmr
11	Z-D-Phe-Pro-BoroPgl-OPinac	41.5%	Nmr
39	Z-D-Dpa-Pro-BoroMpg-OPin		
43	Z-D-Phe-Pro-BoroOct-OPinac	77%	Nmr
51	Z-D-Dpa-Pro-BoroMpg-OPin	quantitative	Nmr

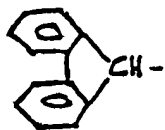
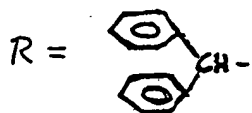
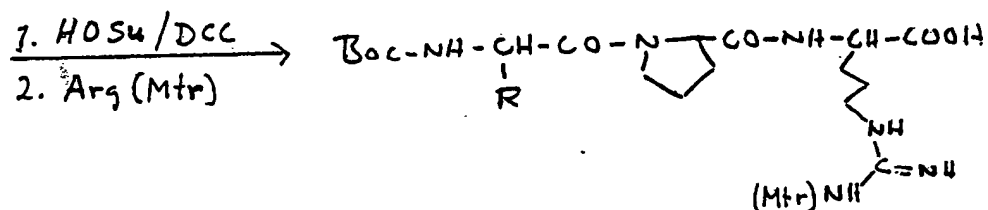
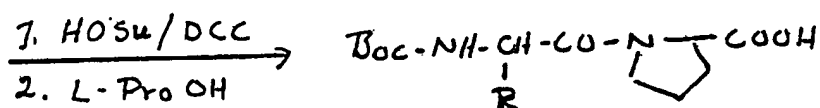
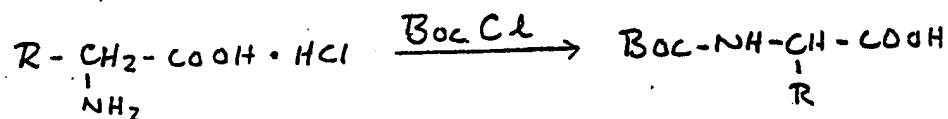
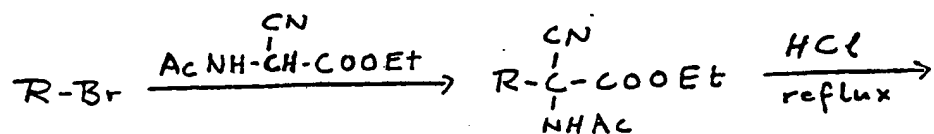
Example 22

The following compounds were synthesised according substantially to Examples 3a and 3b:

12	Z-D-Dpa-Pro-BoroIrg-OPin	61%	Nmr, FABMs[M]780,13%.
26	Z-D-8-Nal-Pro-BoroIrg-OPin	41.8%	Nmr, FABMs[M+H]755,10%.
36	Z-D-Fgl-Pro-BoroIrg-OPin	49%	Nmr, FABMs[M+H]778,11%.
37	Ac-D-Dpa-Pro-BoroIrg-OPin	8.1%	Nmr, FABMs[M+H]689,14%.
38	Z-L-Dpa-Pro-BoroIrg-OPin	39%	Nmr, FABMs[M+H]781,12%.
46	Z-D-Cha-Pro-BoroIrg-OPin	41%	Nmr, FABMs[M+H]711,10%.

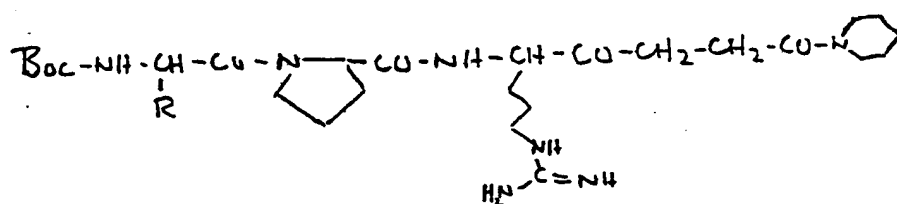
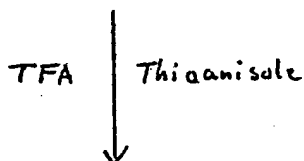
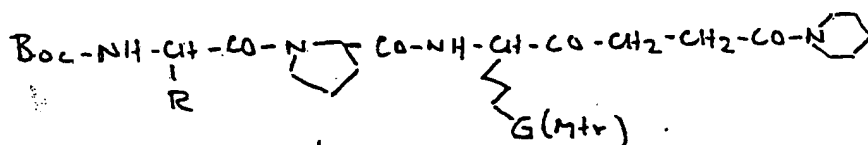
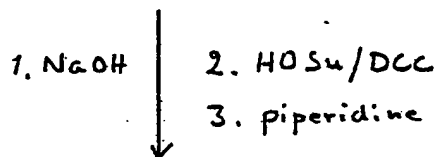
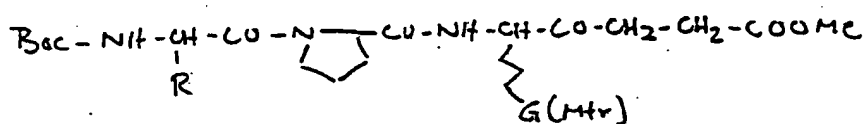
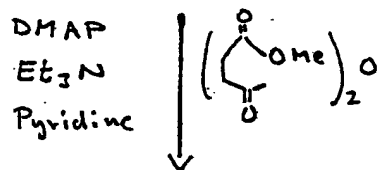
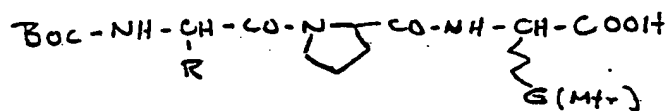
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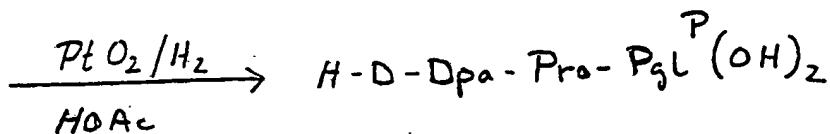
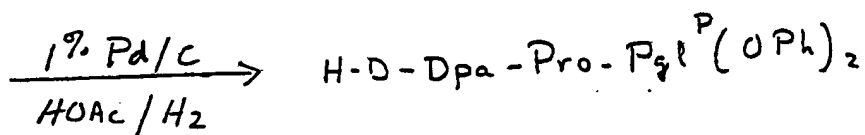
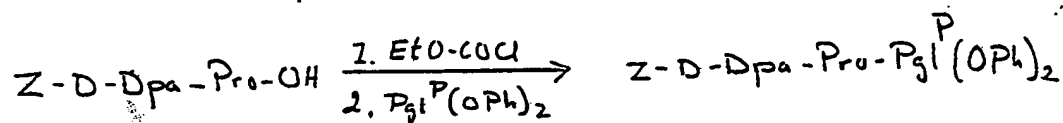
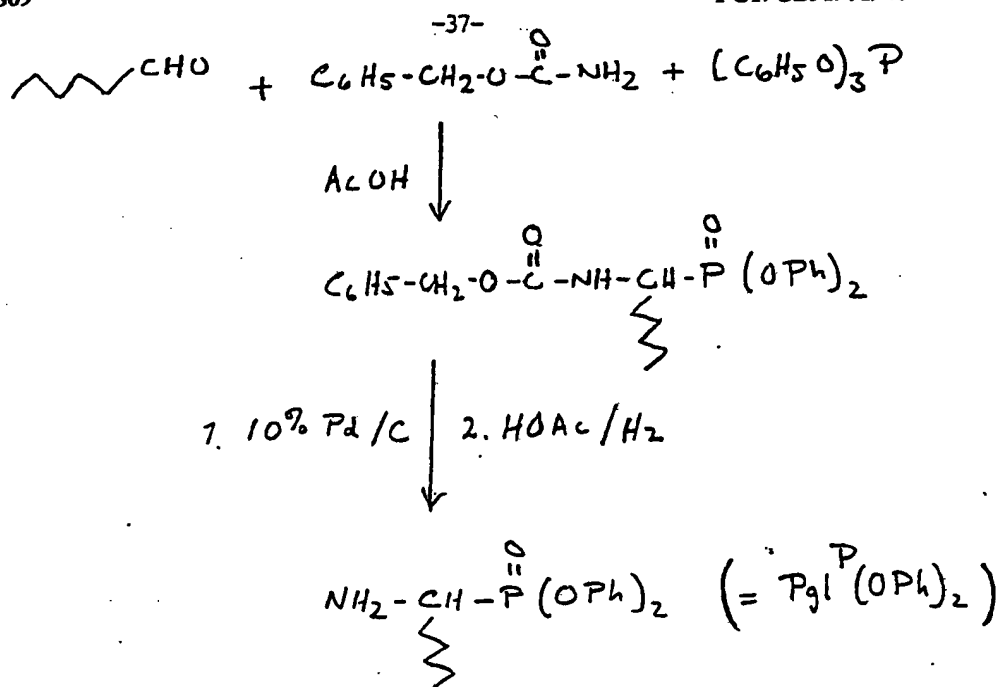
Scheme 1. Synthesis of the amino acids Dpa, Nal and Fgl and their di- and tripeptides with Pro and Pro-Aro

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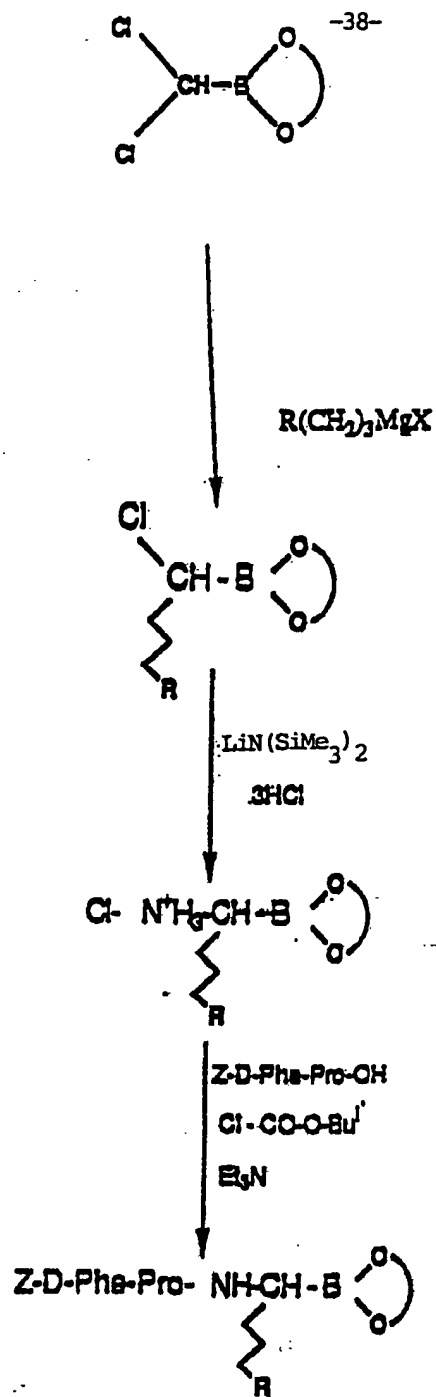


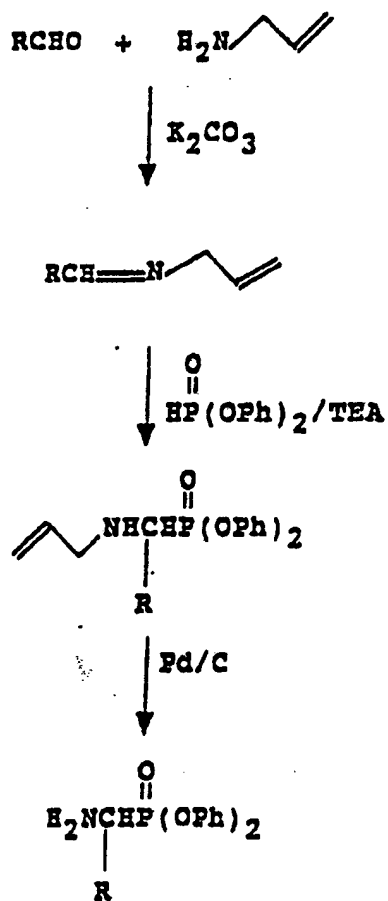
G = guanidino

Scheme 2. Synthesis of ketomethylene isosteric inhibitor

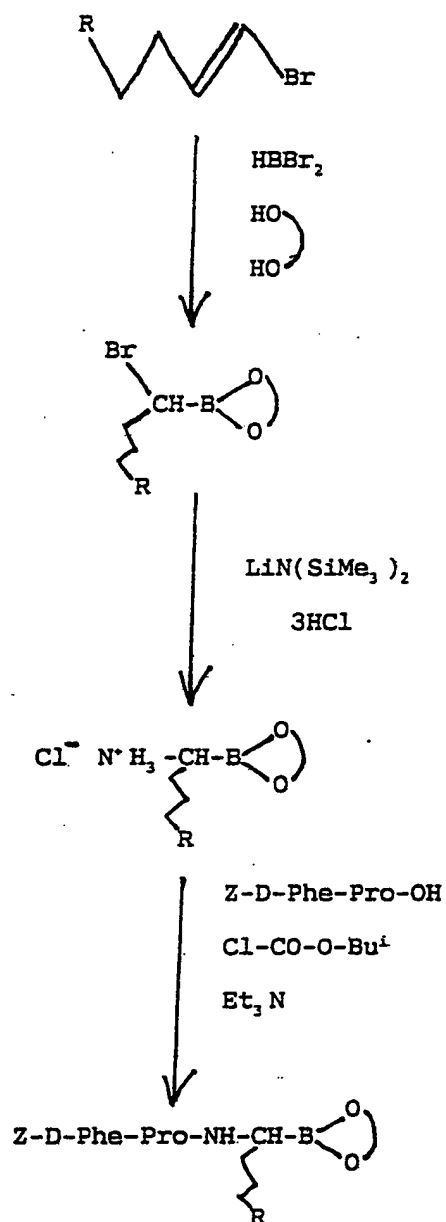


Scheme 3. Synthesis of aminophosphonic acid inhibitors

Scheme 4

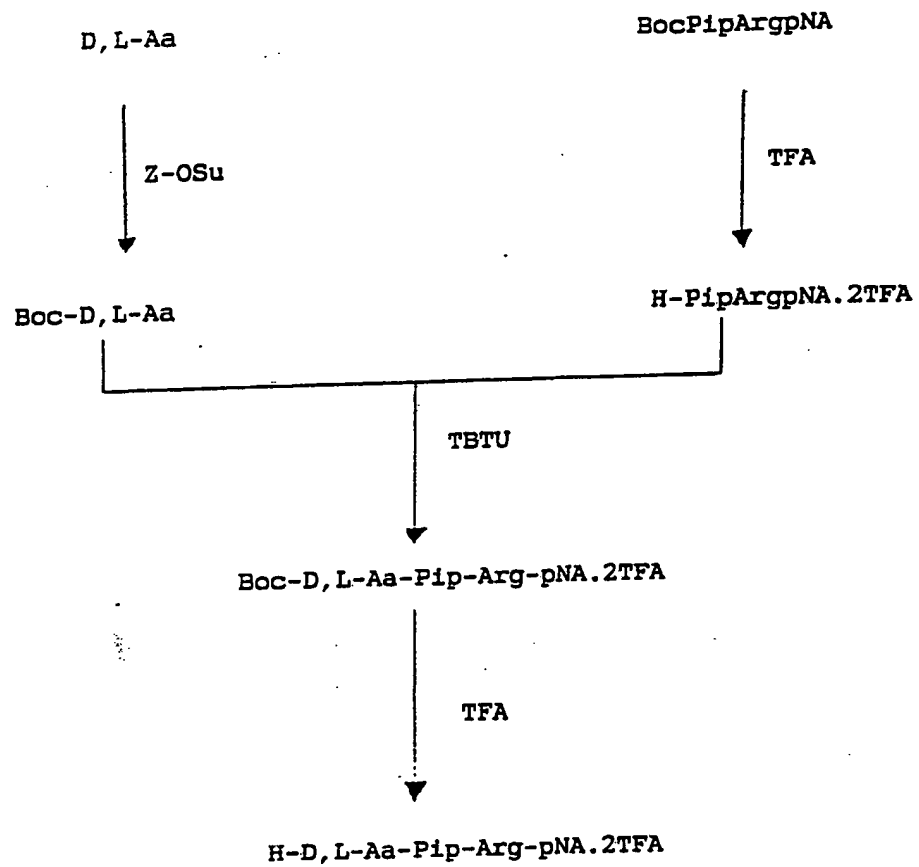
SCHEME 5

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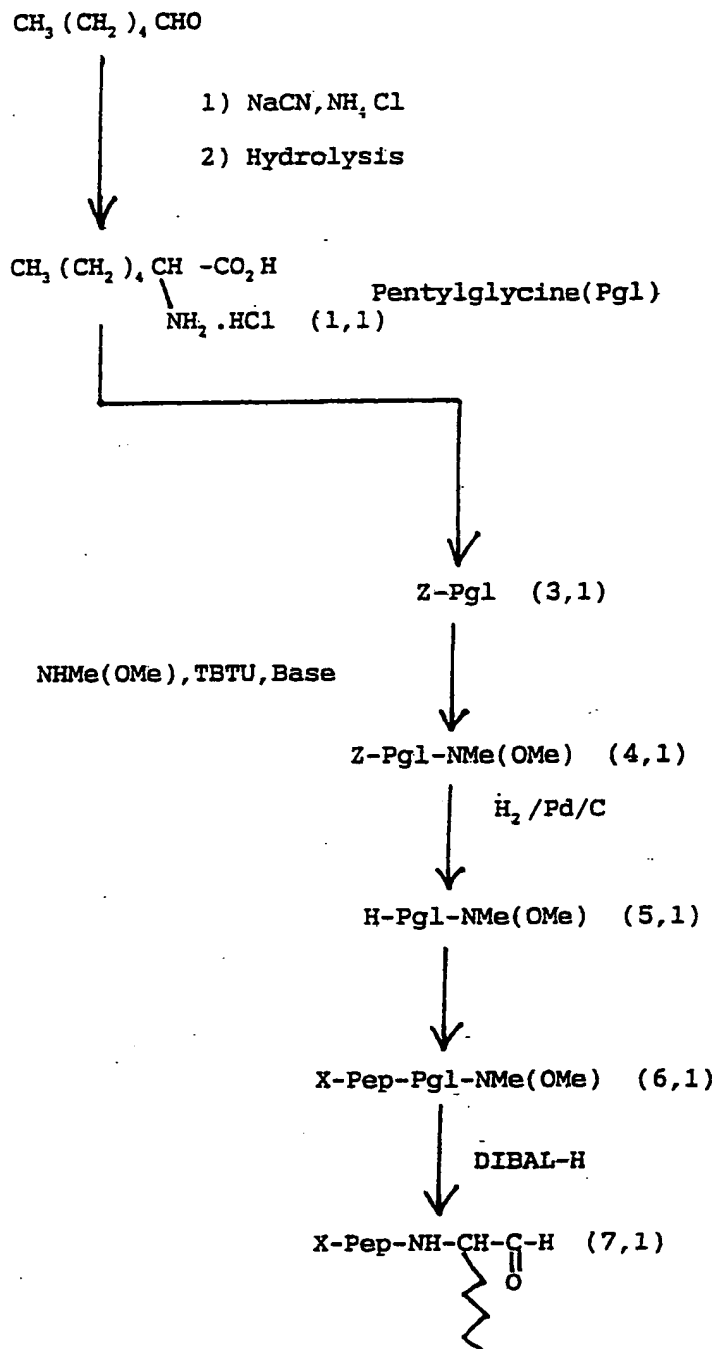
Scheme 6

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Scheme 7

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Scheme 8

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Plasma thrombin time (TT)

A volume of 150µl of citrated normal human plasma and 20µl of buffer or sample were warmed at 37°C for 1 min. Coagulation was started by adding 150µl of freshly prepared bovine thrombin (5NIHu/ml saline) and the coagulation time was recorded on a coagulometer.

A phosphate buffer, pH7.8, containing 0.1% bovine serum albumine and 0.02% sodium azide was used. The samples were dissolved in DMSO and diluted with the buffer. When no inhibitor was used DMSO was added to the buffer to the same concentration as that used in the samples. The inhibitor concentrations were plotted against the thrombin times in a semilogarithmic graph from which the inhibitor concentration that caused a doubling (40 sec) of the thrombin time was determined.

Determination of Ki

The inhibition of human α -thrombin was determined by the inhibition of the enzyme catalyzed hydrolysis of three different concentrations of the chromogenic substrate S-2238.

200µl of sample or buffer and 50µl of S-2238 were incubated at 37°C for 1 min and 50µl of human α -thrombin (0.25 NIHu/ml) was added. The initial rate of inhibited and uninhibited reactions were recorded at 405nm. The increase in optical density was plotted according to the method of Lineweaver and Burke. The K_m and apparent K_m were determined and K_i was calculated using the relationship:

$$K_i = \frac{I}{\frac{K_m \text{ app}}{K_m} - 1}$$

Cardiovascular effects

Cats weighing 2-3kg were anaesthetized with Mebumal, given as an intraperitoneal injection. The central blood pressure and heart rate were recorded on a Grass Polygraph by means of a catheter inserted in the femoral artery.

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DETERMINATION OF k_m

The k_m of substrates with human α -thrombin were determined by measuring the absorbance at a series of dilutions of substrate. (page 753, Longman Scientific & Technical, 5th edition, 1989.)

DETERMINATION OF ACTIVATED PARTIAL THROMBOPLASTIN TIME (APTT) FOR IN-VITRO SAMPLES

A volume of 150 μ l of citrated (3.2%) normal human plasma was incubated at 37°C with sample (20 μ l) or buffer (20 μ l, control) for 1 min. To the solution was added reconstituted "AUTOMATED APTT" (available from Organon Teknika, 0.1ml). Each solution was activated at 37°C for 5 min.

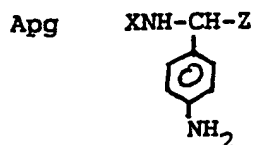
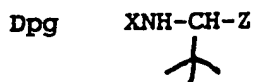
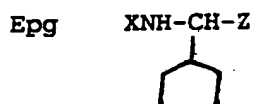
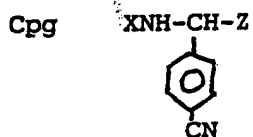
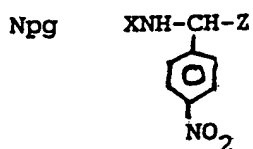
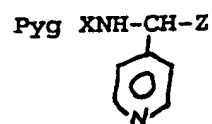
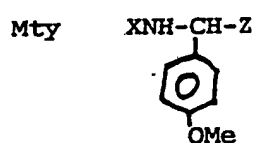
After activation, calcium chloride (0.1ml, 0.025M, prewarmed at 37°C) was added and clot detection was timed using a "semiautomated coagulometer" (Nach, Schnicter und Gross).

IN-VIVO TOXICITY DATA

Deposition of 111 indium-labelled platelets was monitored after 1mg/kg intravenous doses, via a cannula in the marginal ear vein of New Zealand white rabbits, as outlined in G.R.May, C.M.Hero, K.D.BUTLER, C.P.PAGE, JOURNAL OF PHARMACOLOGICAL METHODS, 24, pp1-35, 1990. Peripheral blood pressure was monitored by a pressure sensor in the carotid artery.

EX-VIVO APTT, TT

Data was obtained as for in-vitro tests, but using plasma samples obtained at 0,1,10,30 and 60 minutes after dosing of 1mg/kg, i.v. bolus injection in a marginal ear vein in New Zealand white rabbits, which were anaesthetized from a canula in the carotid artery.



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PLATELET ACCUMULATION

Compound	(% OF NORMAL) LUNG	LEG	BLOOD PRESSURE Δ% of NORMAL
<u>4 mg/kg i.v. DOSE</u>			
No. 25 ZD-Phe Pro Boro ² OPin	FATAL PLATELET ACCUMULATION 100%	FATAL ACCUMULATION 100%	100%
No. 53 BocDPhe Pro His-H	NONE	NONE	0
No. 22 ZD-Phe Pro Boro ² OPin	NONE	11%, TRANSIENT	0
No. 41 ZD-Phe Pro Boro ² PheOPin	NONE	NONE	21%
<u>1 mg/kg i.v. DOSE</u>			
No. 59 ZD-Phe Pro Boro ² OPin	NONE	NONE	NONE
No. 25 ZD-Phe Pro Boro ² OPin	44-40%, PROLONGED	44-40%	SMALL
No. 20 H-D-Phe Pro Arg ¹ (Glu) ₂	NONE	NONE	NONE
No. 53 BocDPhe Pro His-H	NONE	NONE	NONE
No. 6 H-D-Phe Pro Arg ¹ GlyPip	NONE	NONE	NONE

EX-VIVO DATA		T. T.		A. P. T. T.			
COMPOUND NO. 6.							
ANIMAL		1	2	1	2		
TIME (MIN)							
0		24.4	15.6	32.9	27.9	24	57
1		176, 200	260	200	46	78	
10		55	61.4	76	57.5	32	55
30		33	36.9	62	36	34	57
60		25	27.9	49	31.5	27	62

COMPOUND NO. 11.						
		1	2	1	2	
TIME						
0		30	24.5	26	46.5	26.4
1		>300	>200	>300	UC	UC
10		>300	>200	>300	UC	UC
30		102	>200	>300	236.7	49
60		94	>200	>300	53.0	38.4

COMPOUND NO. 59									
TIME (MIN)									
0		28.9	22.5	22.4	23.5	42	59.5	43.9	58
1		111	106	171.5	168.5	111.4	111	102.9	126
10		35.9	34	37.9	38.4	48.9	53	60.5	67
30		29	30	32	31	34.9	49	59	53.9
60		21.4	28.5	27.9	28	40.4	48.4	65.5	62

		T.T.		A.P.T.T.			
ANIMAL		1	2	1	2		
COMPOUND NO.	53.						
TIME (MIN)							
0	26	27	24	24.9	32.9	24.9	
1	86.9	83.5	80.9	74	41.5	41.5	27.9 30.9
10	32	32	30	29.9	39.9	29.9	26 25.9
30	24.4	25	25.9	25.4	31.4	30.4	25.4 26.1
60	24.4	23.9	22.9	22.9	28.9	27.5	25.9

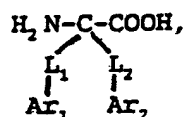
	(C50 No incubation)	Inc.	Insulin no inc / inc	Xa no inc / inc	Urokinase no inc / inc	Kallikrein no inc / inc	Cytomolydin no inc / inc
15 Boc-Dpa-Pro-IgP(OH) ₂	5.28	0.0836					
16 D,L-Dpa-Pro-IgP(OH) ₂	5.16	0.0817					
18 D-Dpa-Pro-IgP(OH) ₂	148.0	0.151					
19 Z-D-Phe-Pro-IgP(OH) ₂	149.0	0.0187					
21 D-Phe-Pro-IgP(OH) ₂	nom(137.0)	4.35					
23 D-Dpa-Pro-IgP(OH) ₂	0.187	0.00938					
31 H-D-Dpa-Pro-MgP(OH) ₂	1.48	0.00235	n(187.0)/332	39.7/n18.7	83.0/n18.7	66.7/n18.7	n18.7/n0.000149
40 H-D-Phe-Pro-MgP(CPh) ₂	16.6	0.0118	n20.9 /52.5	n20.9 /n20.9	n20.9 /n20.9	n20.9 /n20.9	n20.9 /0.00263
55 H-D-Phe-Pro-ArgP(OH) ₂		3.96					
56 H-D-Phe-Pro-ArgP(OH) ₂		0.417					
57 H-D-Phe-Pro-DPpP(OH) ₂		0.0828					
22 Z-D-Phe-Pro-BornMgP(OH) ₂	0.007		11.5	3.86	9.13	12.9	
41 Z-D-Phe-Pro-BornMgP(OH) ₂	0.0128		37.3	7.94	6.67	16.7	
50 Z-D-Phe-Pro-BornMgP(OH) ₂	0.0007		3.23	1.72	3.63	7.26	
61 D-Dpa-Pro-Arg-k-Gly-Pip	0.2		n(157.0)	149.0	494.0	199.0	
44 D,L-Phe-Pro-BornMgP(OH) ₂	1.3		0.153	0.512	0.121	1.08	
25 Z-D-Phe-Pro-BornMgP(OH) ₂	1.5	0.068	0.0084	0.0141	0.075	0.334	
52 Boc-D-Phe-Pro-Ile-Ile			n(413.0)	n(413.0)	n(413.0)	nom(413.0)	

References

1. Claeson, G. and Aurell, L. Annals of New York Academy of Sciences, 370, 79-811 (1981).
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3. Kettner, C. and Shaw, E. Thrombosis Research 14, 969-973 (1979).
4. Szelke, M. and Jones, D.M. US patent 4,638,047-A.
5. Kettner, C. and Shenvi, A.B. European patent appl. 293881 (1988).
6. Stüber, W., Kosina, H. and Heimbürger, N. Int. J. Peptide Protein Res. 31, 63-70 (1988).
7. Kaiser, B., Hauptmann, J. and Markwardt, F. Die Pharmacie 42, 119-121 (1987).

CLAIMS

1. A peptide derived from the formula
D-Phe-Pro-Arg
or its analogues, wherein Phe is substituted by



wherein

Ar_1 and Ar_2 are the same or different and are selected from the group consisting of phenyl, thienyl, pyridyl, naphthyl, thionaphthyl, indolyl and saturated groups corresponding to these, optionally substituted by up to three groups selected from C_1 - C_3 alkyl and C_1 - C_3 alkoxy,

L_1 and L_2 are the same or different and are selected from the group consisting of CH_2 , CH_2 - CH_2 , $\text{O}-\text{CH}_2$, $\text{S}-\text{CH}_2$,

$\text{Ar}-\text{L}$ taken together L ^{optionally} means H, diphenyl-methyl, fluorenyl or saturated groups corresponding to these, but one of the $\text{Ar}-\text{L}$ cannot be H when the other $\text{Ar}-\text{L}$ means H or benzyl.

2. A peptide according to claim 1, wherein Phe is substituted by Dpa, Nal or Dba.

3. A peptide according to claim 1, wherein Arg is substituted by $\text{H}_2\text{N}-\text{CH}-\text{COOH}$,



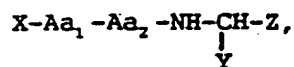
wherein

$\text{Y} = [\text{CH}_2]_n-\text{Q}$, $\text{CH}_2-\text{C}_6\text{H}_4-\text{Q}$ where $\text{Q} = \text{H}$, amino, amidino, imidazole, guanidino or isothioureido and $n = 1-5$, preferably 3-5, or C_3 - C_9 alkyl and C_5 - C_{10} aryl or alkylaryl optionally substituted by up to three groups selected from hydroxy and C_1 - C_3 alkoxy.

4. A peptide according to claim 3, wherein Arg is substituted by Irg, Gpa, Apa or a non-basic amino acid.


5. A peptide according to claim 4, wherein the non-basic amino acid is selected from Pgl, Mbg and Chg.

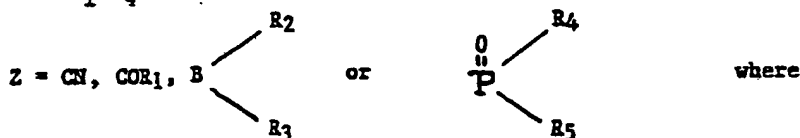
6. A peptide of the formula



wherein

X = H, CH₃ or an N-protecting group, e.g. Ac, Bz, Chz, Boc;

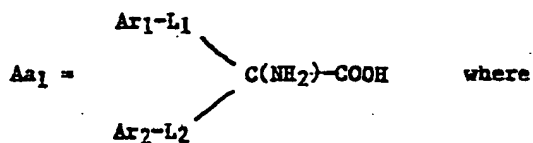
Y = [CH₂]_n-Q, CH₂--Q where Q = H, amino, amidino, imidazole, guanidino or isothioureido and n = 1-5, preferably 3-5, or C₃-C₉ alkyl and C₅-C₁₀ aryl or alkylaryl optionally substituted by up to three groups selected from hydroxy and C₁-C₄ alkoxy;



R₁ = H, OH, CH₂Cl, CH₂-CH₂-CO-pip, CF₂-CF₂-CO-pip, CH₂-CH(CO-pip)-CH₃, CF₂-CF(CO-pip)-CF₃,
CH₂-CH₂-CO-Pro-NHET, CF₂-CF₂-CO-Pro-NHET or a chromophoric group e.g. pNA, MCA,

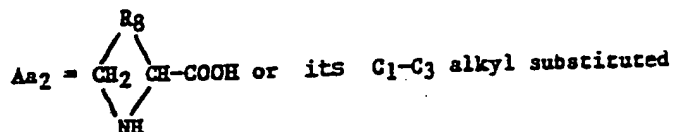
R₂ and R₃ are the same or different and are selected from the group consisting of OH, OR₆ and NR₆R₇, or R₂ and R₃ taken together represent the residue of a diol; where R₆ and R₇, which are the same or different, are C₁-C₁₀ alkyl, phenyl or C₆-C₁₀ arylalkyl,

R₄ and R₅ are the same or different and are selected from R₂, R₃, Gly-pip, Ala-pip or Gly-Pro-NHET;



Ar₁ and Ar₂ are the same or different and are selected from the group consisting of phenyl, thienyl, pyridyl, naphthyl, thionaphthyl, indolyl and saturated groups corresponding to these, optionally substituted by up to three groups selected from C₁-C₃ alkyl and C₁-C₃ alkoxy,

L_1 and L_2 are the same or different and are selected from the group consisting of CH_2 , CH_2-CH_2 , $O-CH_2$, $S-CH_2$,
 $Ar-L$ taken together ^{optionally} means H, diphenyl-methyl, fluorenyl or saturated groups corresponding to these, but one of the $Ar-L$ cannot be H when the other $Ar-L$ means H or benzyl;



derivatives, where $R_3 = CH_2$, CH_2-CH_2 , $S-CH_2$, $S-C(CH_3)_2$ or $CH_2-CH_2-CH_2$.

7. A peptide according to claim 6 which is selected from the following:

Ac-D- β Nal-Pro-boroArg pinanediol ester

Z-D-Dpa-Pro-boroIrg pinanediol ester

Z-D-Dpa-Pro-boroPgl pinacol ester

Ac-D- β Nal-Pro-boroMbg pinanediol ester

CH_3 -D-Dpa-Pro-Arg-H

Boc-D-Dpa-Pro-Gpa-H

CH_3 -D-Dpa-Thi-Mbg-H

H-D-Dpa-Pro-Arg-k-Gly-pip

Z-D-Dpa-Pro-Arg- CH_2Cl

Boc-D-Dpa-Pro-ArgCN

H-Dpa-Pro-Arg^P (OPh)₂

H-D- β Nal-Pro-Pgl^P (OPh)-Gly-pip

H-D-Dpa-Pip-Arg-pNA

H-D- β Nal-Pro-Chg-pNA

and the compounds listed in Examples 10 to 22 herein.

8. A substrate of thrombin, which is or includes a peptide according to any one of claims 1 to 7.
9. An inhibitor of thrombin, which is or includes a peptide according to any one of claims 1 to 7.
10. A method of inhibiting thrombin in a mammalian host, comprising administering an effective amount of a peptide according to any one of claims 1 to 7.
11. A method according to claim 10, wherein the peptide is administered in combination with a pharmaceutically acceptable carrier or diluent.
12. A method according to claim 10, wherein the peptide is administered at a dose of from 0.02 to 15mg/kg of body weight.
13. A method according to claim 12, wherein the dose is from 1 to 10mg/kg of body weight.
14. A method according to claim 10, wherein the peptide is administered as a single dose or in divided doses or as a sustained release formulation.
15. A method according to claim 10, wherein the host is a human host.
16. A method of preventing coagulation of mammalian blood, comprising adding thereto an effective amount of a peptide according to any one of claims 1 to 7.
17. A method according to claim 16, wherein the amount of peptide added of from 1 to 10mg/litre.
18. A method of establishing an extracorporeal blood loop for a patient, comprising administering intravenously from 0.1 to 1mg/kg of body weight of a peptide according to any one of claims 1 to 7.

19. Use of a peptide according to any one of claims 1 to 7 as an inhibitor or substrate of thrombin.

20. Use of a peptide according to any one of claims 1 to 7 in the preparation of a medicament for the treatment or prevention of thrombosis.

21. A pharmaceutical composition for inhibiting thrombin in a mammalian host, comprising a peptide according to any one of claims 1 to 7 and a pharmaceutically acceptable carrier or diluent.

INTERNATIONAL SEARCH REPORT

International Application No PCT/GB 91/01946

I. CLASSIFICATION OF SUBJECT MATTER (If several classification symbols apply, indicate all) ⁶		
According to International Patent Classification (IPC) or to both National Classification and IPC		
IPC5: C 07 K 5/08, 5/06, A 61 K 37/64, C 12 Q 1/56		
II. FIELDS SEARCHED		
Minimum Documentation Searched ⁷		
Classification System	Classification Symbols	
IPC5	A 61 K; C 07 K; C 12 Q	
Documentation Searched other than Minimum Documentation to the Extent that such Documents are Included in Fields Searched ⁸		
III. DOCUMENTS CONSIDERED TO BE RELEVANT⁹		
Category [*]	Citation of Document, ¹¹ with indication, where appropriate, of the relevant passages ¹²	Relevant to Claim No. ¹³
X	US, A, 4428874 (LARS G. SVENDSEN) 31 January 1984, see the whole document --	1,3,6,8, 19
X	US, A, 4450105 (TAKESHI NAGASAWA ET AL) 22 May 1984, see the whole document --	1,3,6,8, 19
A	US, A, 4929602 (LAURENCE A. HARKER ET AL) 29 May 1990, see the whole document --	1,3,6,9, 19-21
<p>[*] Special categories of cited documents:¹⁰</p> <p>"A" document defining the general state of the art which is not considered to be of particular relevance</p> <p>"E" earlier document but published on or after the international filing date</p> <p>"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)</p> <p>"O" document referring to an oral disclosure, use, exhibition or other means</p> <p>"P" document published prior to the international filing date but later than the priority date claimed</p> <p>"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention</p> <p>"X" document of particular relevance, the claimed invention cannot be considered novel or cannot be considered to involve an inventive step</p> <p>"Y" document of particular relevance, the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art.</p> <p>"&" document member of the same patent family</p>		
IV. CERTIFICATION		
Date of the Actual Completion of the International Search	Date of Mailing of this International Search Report	
10th February 1992	21 FEB 1992	
International Searching Authority	Signature of Authorized Officer	
EUROPEAN PATENT OFFICE	T. TAZELAAK	

III. DOCUMENTS CONSIDERED TO BE RELEVANT (CONTINUED FROM THE SECOND SHEET)		
Category *	Citation of Document, with indication, where appropriate, of the relevant passages	Relevant to Claim No
A	EP, A2, 0235692 (BEHRINGWERKE AG) 9 September 1987, see the whole document --	1,3,6,9, 19-21
A	WO, A1, 8909612 (CORVAS, INC.) 19 October 1989, see the whole document -- -----	1,3,6,9, 19-21

FURTHER INFORMATION CONTINUED FROM THE SECOND SHEET

V. ☒ OBSERVATIONS WHERE CERTAIN CLAIMS WERE FOUND UNSEARCHABLE¹

This international search report has not been established in respect of certain claims under Article 17(2) (a) for the following reasons:

1. ☒ Claim numbers 10-15 because they relate to subject matter not required to be searched by this Authority, namely:

Method for treatment of the human or animal body by therapy. Rule 39(iv).

2. ☐ Claim numbers..... because they relate to parts of the international application that do not comply with the prescribed requirements to such an extent that no meaningful international search can be carried out, specifically:

3. ☐ Claim numbers..... because they are dependent claims and are not drafted in accordance with the second and third sentences of PCT Rule 6.4(a).

VI. ☐ OBSERVATIONS WHERE UNITY OF INVENTION IS LACKING²

This International Searching Authority found multiple inventions in this international application as follows:

1. ☐ As all required additional search fees were timely paid by the applicant, this international search report covers all searchable claims of the international application.

2. ☐ As only some of the required additional search fees were timely paid by the applicant, this international search report covers only those claims of the international application for which fees were paid, specifically claims:

3. ☐ No required additional search fees were timely paid by the applicant. Consequently, this international search report is restricted to the invention first mentioned in the claims. It is covered by claim numbers:

4. ☐ As all searchable claims could be searched without effort justifying an additional fee, the International Searching Authority did not invite payment of any additional fee.

Remark on Protest

- ☐ The additional search fees were accompanied by applicant's protest.
☐ No protest accompanied the payment of additional search fees.

**ANNEX TO THE INTERNATIONAL SEARCH REPORT
ON INTERNATIONAL PATENT APPLICATION NO. PCT/GB 91/01946**

SA 52917

This annex lists the patent family members relating to the patent documents cited in the above-mentioned international search report.
The members are as contained in the European Patent Office EDP file on 31/10/91
The European Patent office is in no way liable for these particulars which are merely given for the purpose of information.

Patent document cited in search report	Publication date	Patent family member(s)	Publication date
US-A- 4428874	31/01/84	US-A- 4568636	04/02/86
US-A- 4450105	22/05/84	EP-A-B- 0076042	06/04/83
		JP-C- 1574613	20/08/90
		JP-A- 58056695	04/04/83
		US-A- 4520100	28/05/85
US-A- 4929602	29/05/90	AU-B- 599280	12/07/90
		AU-D- 3286489	05/07/89
		EP-A- 0343241	29/11/89
		WO-A- 89/05148	15/06/89
EP-A2- 0235692	09/09/87	AU-B- 611850	27/06/91
		AU-D- 6954887	03/09/87
		DE-A- 3606480	03/09/87
		JP-A- 62207251	11/09/87
		US-A- 4927809	22/05/90
WO-A1- 8909612	19/10/89	AU-B- 617169	21/11/91
		AU-D- 3413589	03/11/89
		EP-A- 0364561	25/04/90
		JP-T- 3502578	13/06/91
		US-A- 5023236	11/06/91

For more details about this annex : see Official Journal of the European patent Office, No. 12/82